

Phase and index imaging for biologie

Pierre Bon – Olivier Haeberlé



MiFoBio 2021

Presqu'île de Giens - France, November 5-12, 2021

Intensity Microscopy

Dark-field

Oblique illumination

Rheinberg illumination

Hoffman modulation

Polarized microscopy

Phase contrast (Zernike)

Differential Phase Contrast (Nomarski)

Ultramicroscope (Siedentopf & Zsigmondy)

- :-) Some control of the illumination**
- :-) A certain comprehension of the light specimen interaction phenomenon**
- :-(Inherently 2-D**

Phase Microscopy

:-(Usually, one records intensity only images

Gabor Holography

Phase Stepping Holography

Off-Axis Holography

Front Wave Analyser

=> One can (now easily) measure amplitude and phase of the diffracted field

Commercial DHM



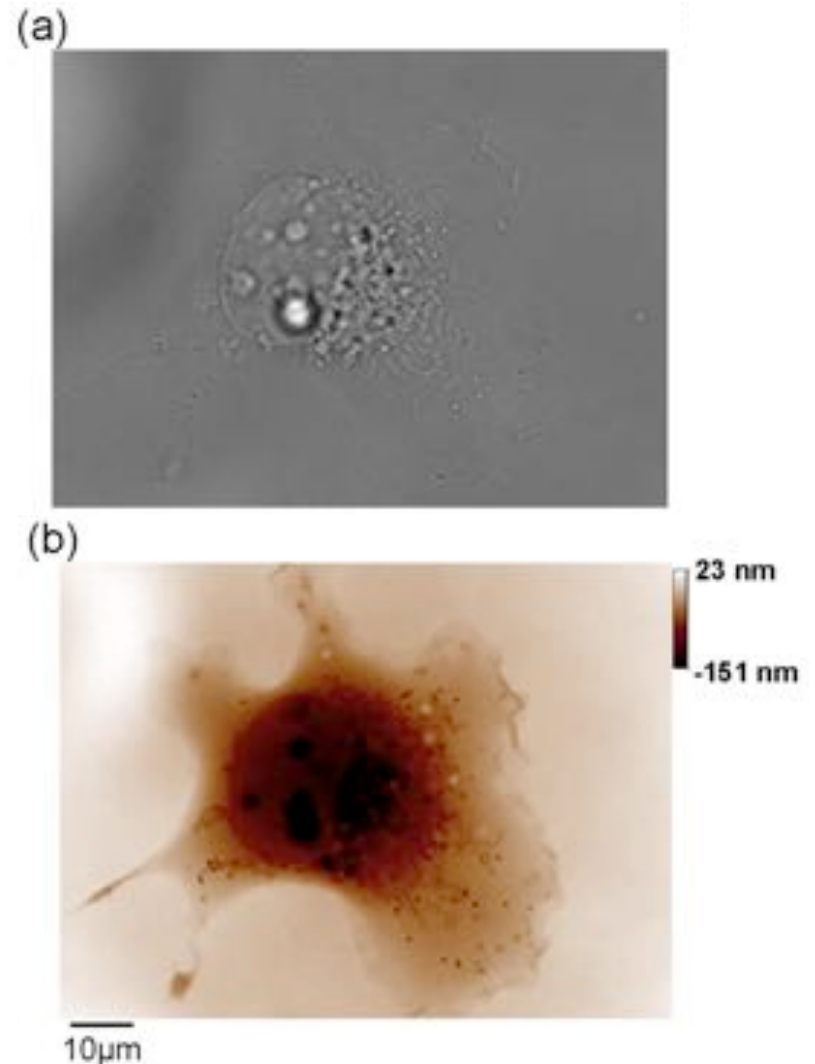
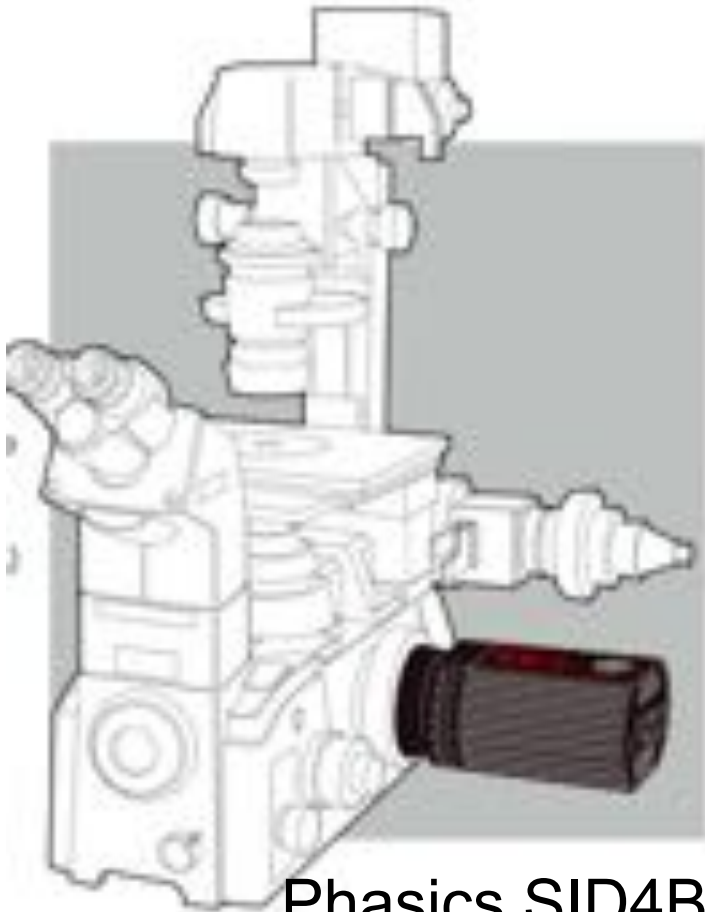
 Lyncée tec ^{DM}



Phase Holographic Imaging

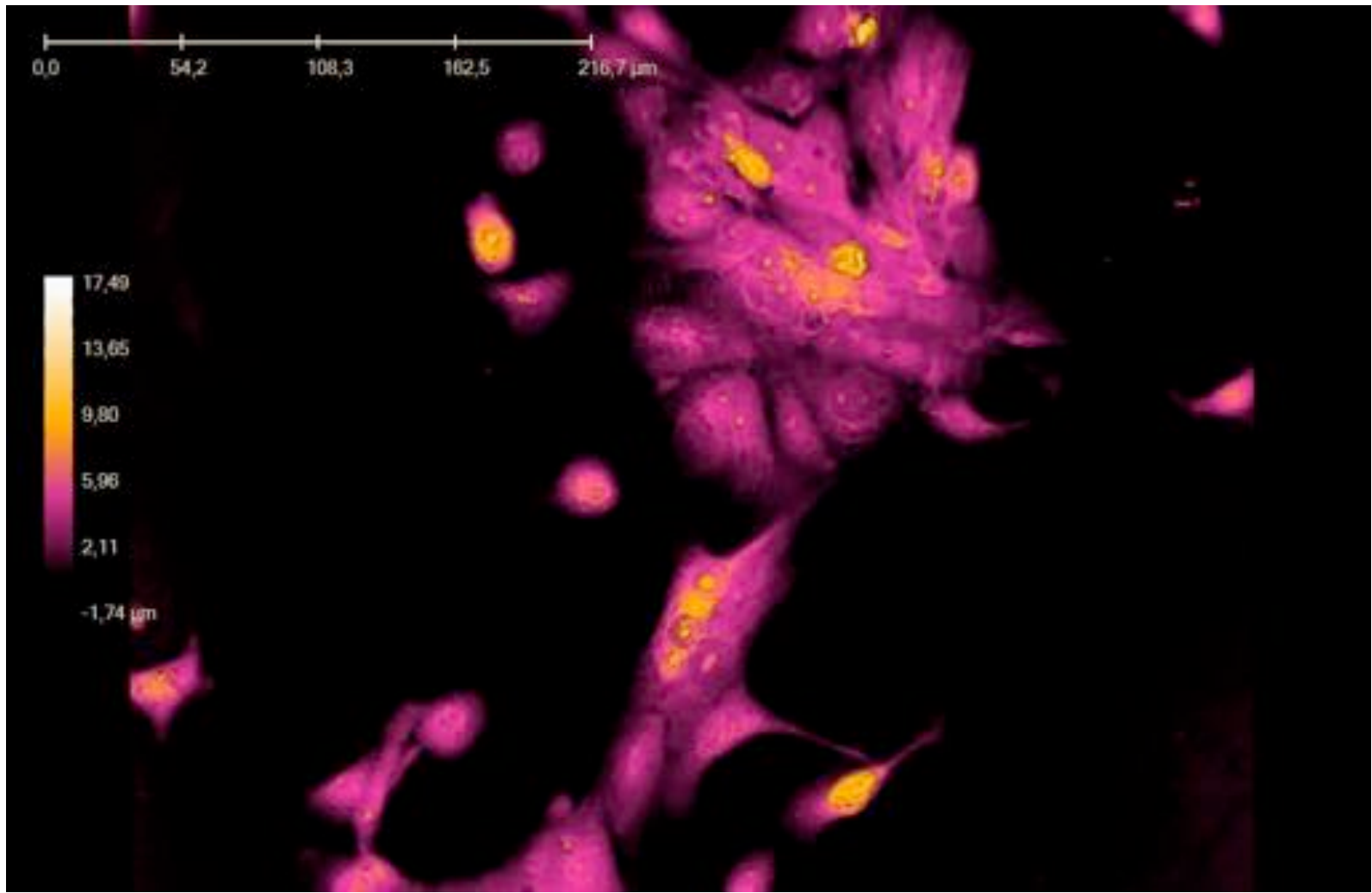


Wavefront Analyser



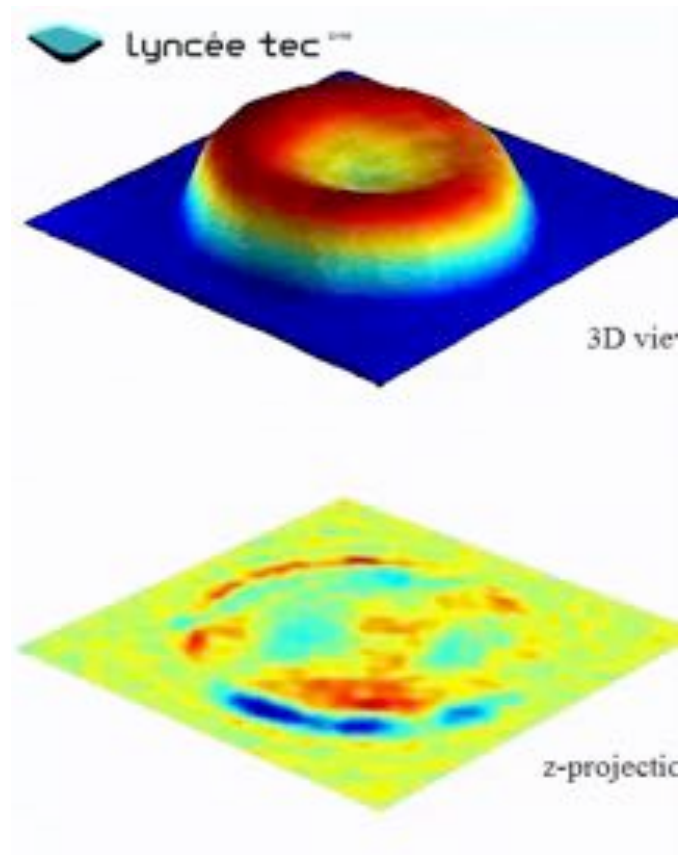
P. Bon, et al., Opt. Expr. **17**, p. 13080 (2009)

Suivi de cultures (DHM)



**Living JIMT-1 cells, recording over 72 hours, 1 frame / 4 min
Birgit Janicke – Phase Holographic Imaging**

Holographic Microscopy

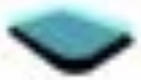


Measurement of the integral refractive index and dynamic cell morphometry of living cells with digital holographic microscopy

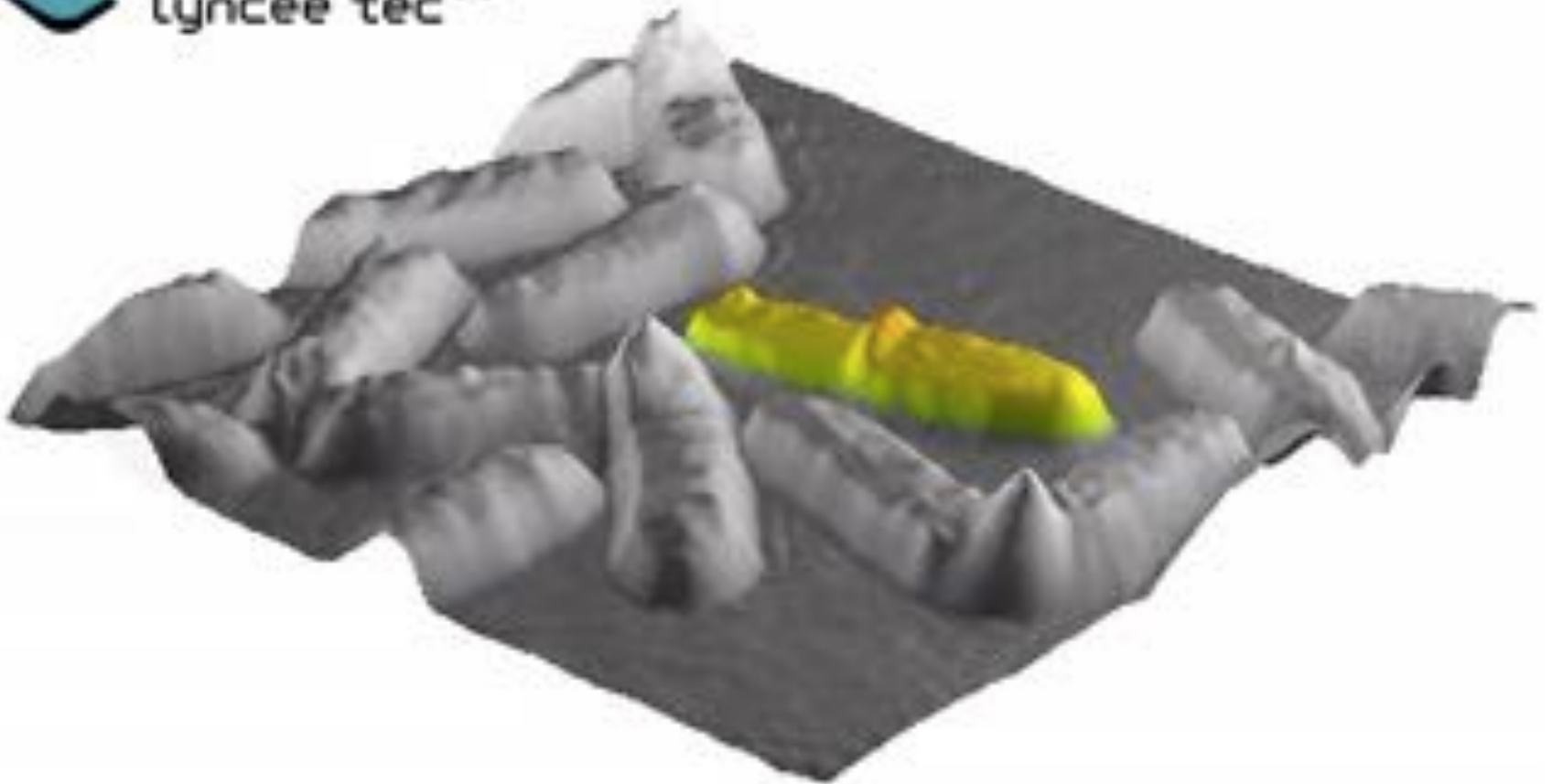
B. Rappaz, *et al.*,

Opt. Express 13 (23), 9361-9373 (2005)

Cartographie de surface



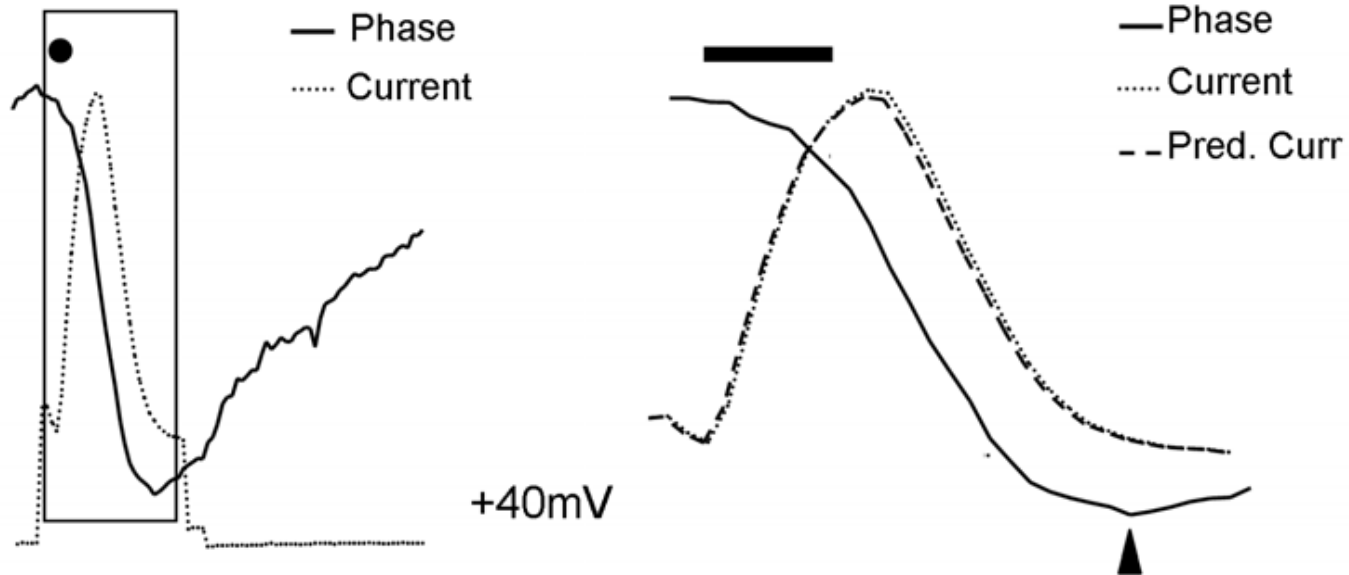
Lyncée tec™



Electrophysiological Optique (DHM)

$$I_{GABA}(t) = \frac{V_0}{\varepsilon_{GABA}^*} \frac{d}{dt} \left(\frac{\varphi_0}{\varphi(t)} \right)^{1/s-r}$$

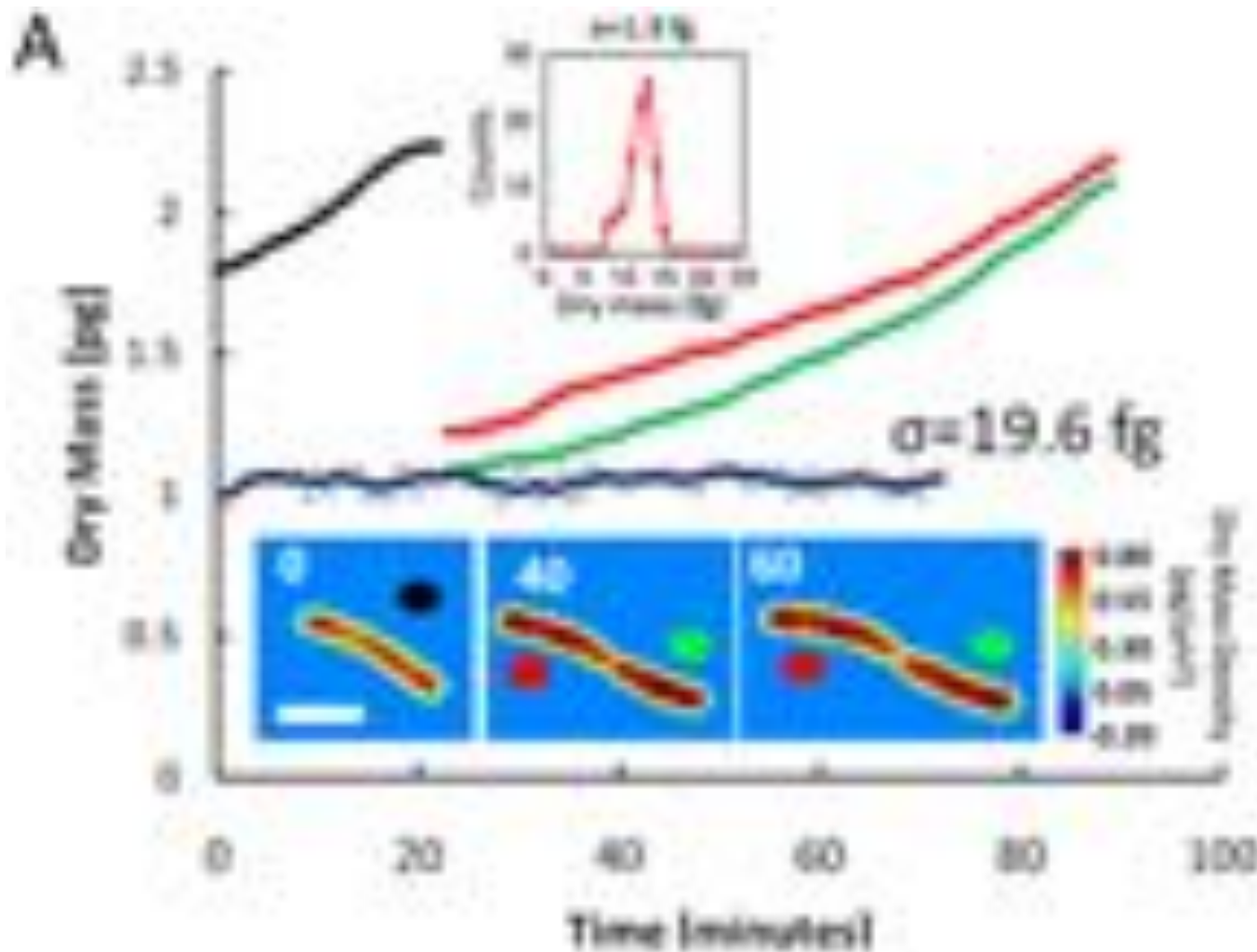
NaCl



"Simultaneous Optical Recording in Multiple Cells by Digital Holographic Microscopy of Chloride Current Associated to Activation of the Ligand-Gated Chloride Channel GABAA Receptor"

P. Jourdain, *et al.*, Plos One 7, e51041(2012)

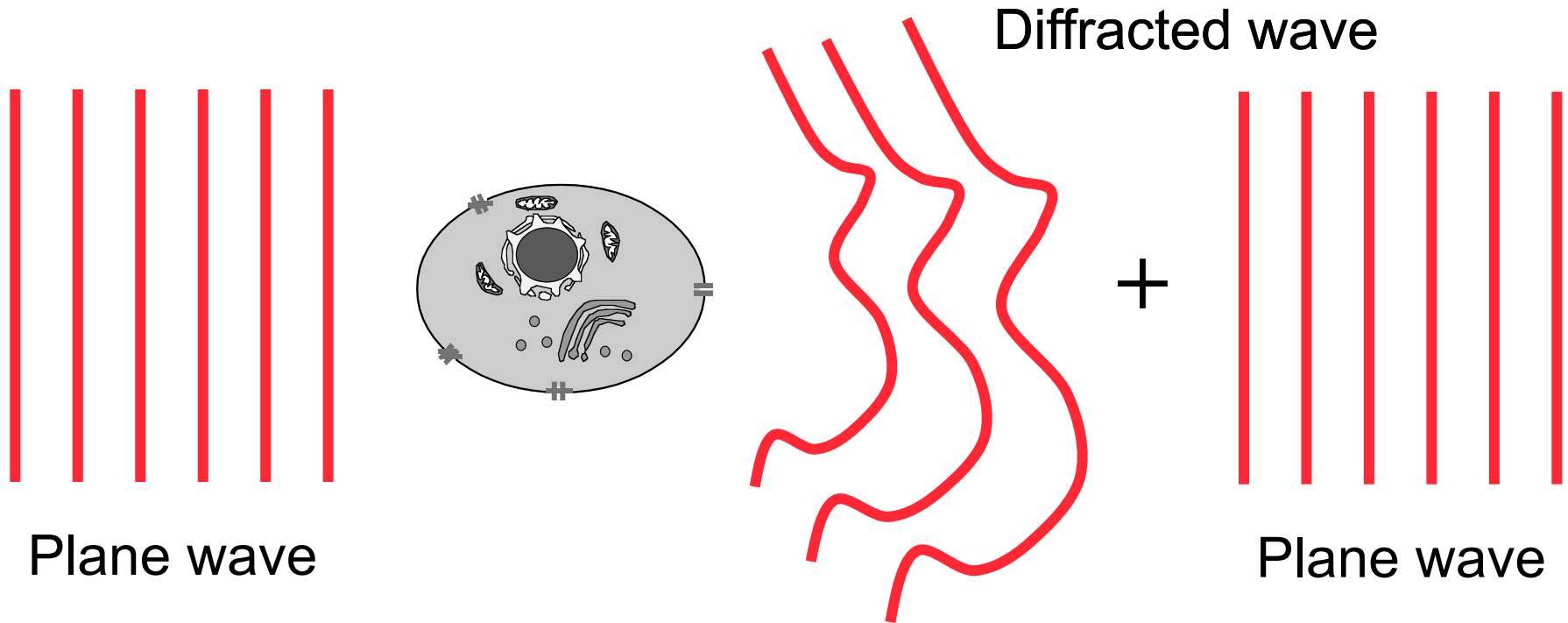
Mesure de masse sèche (SLIM)



“Optical measurement of cycle-dependent cell growth”

M. Mir, *et al.*, PNAS 108, p. 13124 (2011)

Coherent Light Diffraction



Weakly diffracting/diffusing/absorbing object

1st Born approximation

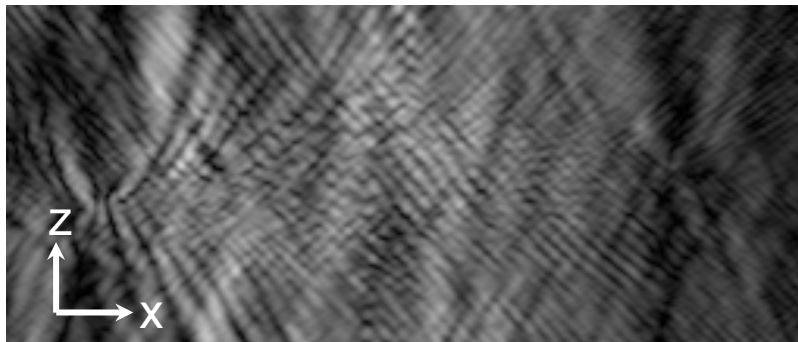
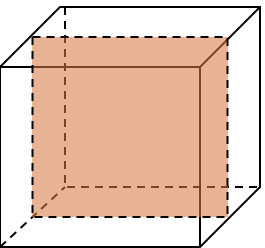
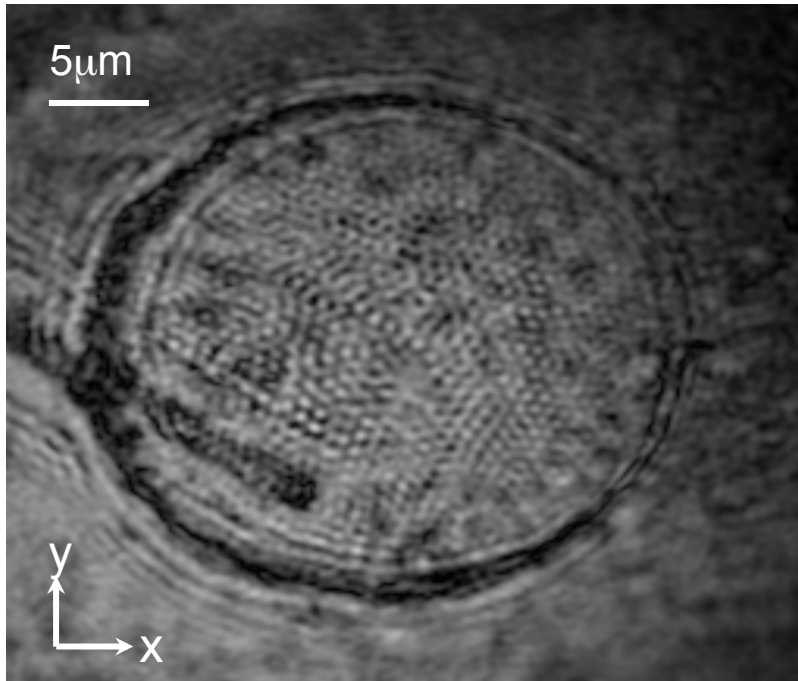
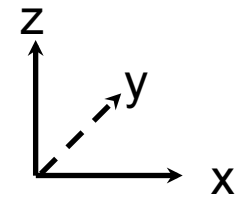
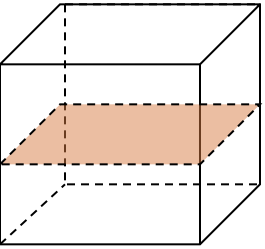
The diffracted wave is interpreted as a part of the 3-D Fourier 3D transform of $\langle n \rangle$

Semi-transparent object **reconstruction from holographic data**

E. Wolf, Opt. Comm. **1**, p. 153 (1969)

Holographic Microscopy: Results

1 angle

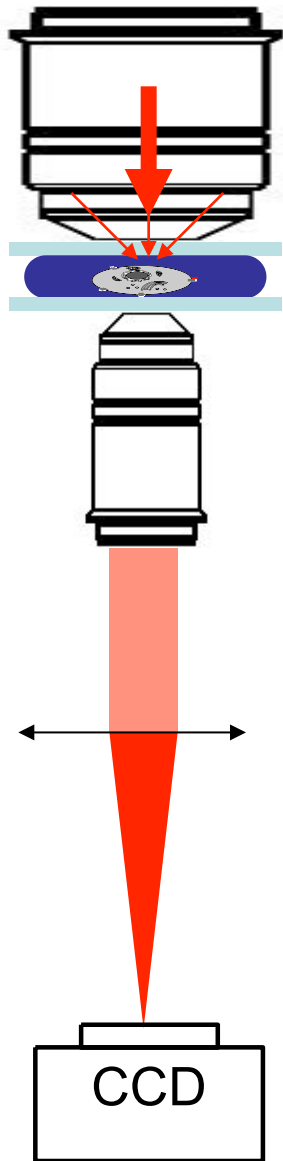


Limited 3-D resolution

Profilometry

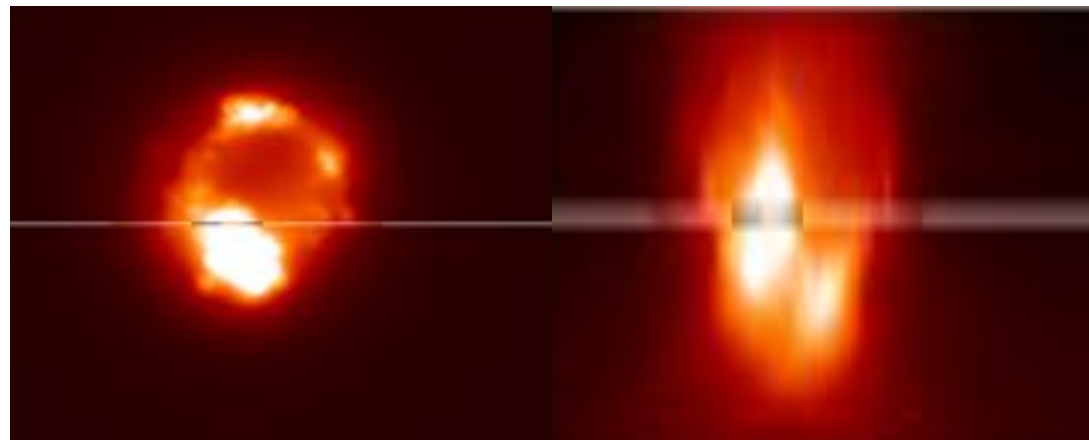
Integral measurements

Illumination Control: Consequences



A radical solution:

1 unique illumination direction!



Cellule CD34 - Image Georges Jung, Laboratoire d'Hématologie
Centre Hospitalier Régional Emile Muller - Mulhouse

$$R_{trans} = R_{fluo} = \frac{\lambda}{(NA_{cond} NA_{obj})}$$

Limits of Label-free VS fluorescence

Sensitivity

→ Not a single molecule approach...

Resolution limit

→ Initial resolution = 50% of fluo. resolution

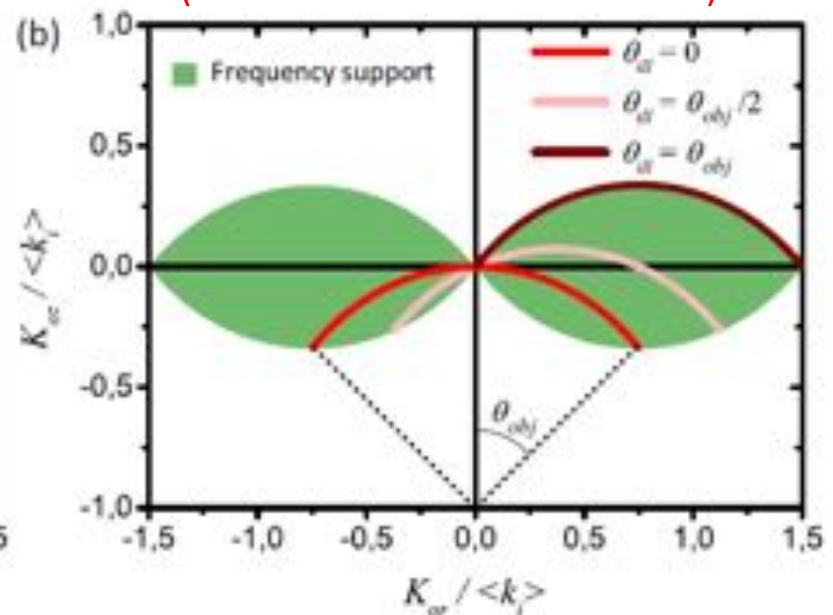
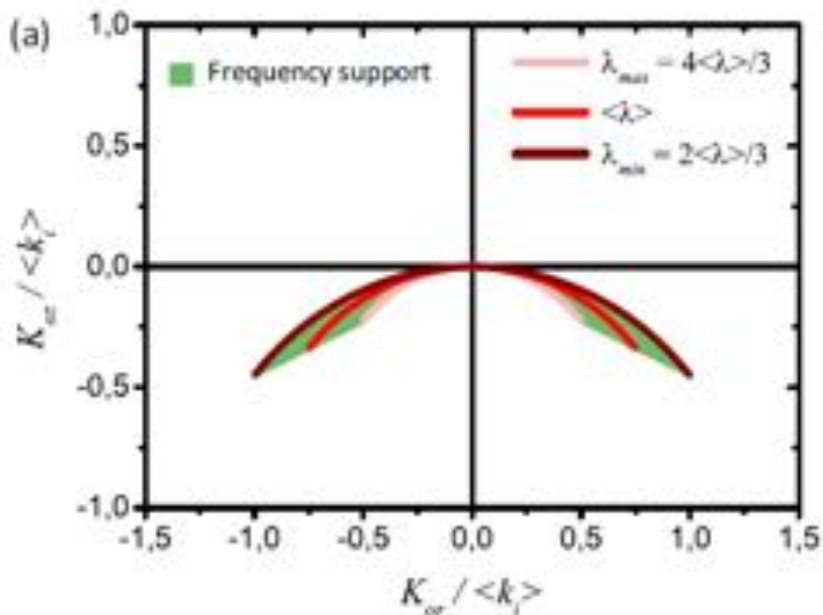
Selectivity/Specificity

→ How to identify and contrast a molecular type

Resolution limit

- Label-free = coherent process VS Fluo = incoherent process
 - Phase imaging = usually interferometric → One illumination angle
 - PSF phase imaging = $1,22\lambda/NA$ VS PSF fluo. = $1,22\lambda/2NA$

(Worst in the axial direction!!!!)



Resolution enhancement

- Aperture synthesis → Multiplexing illumination angles

Bertrand *et al.*, High-resolution tomographic diffractive microscopy of biological samples, **J. Biophotonics**, 3(7), 2010

- Incoherent illumination

Parthasarathy *et al.*, Quantitative phase imaging using a partitioned detection aperture, **Opt. Express**, 37(19), 2012

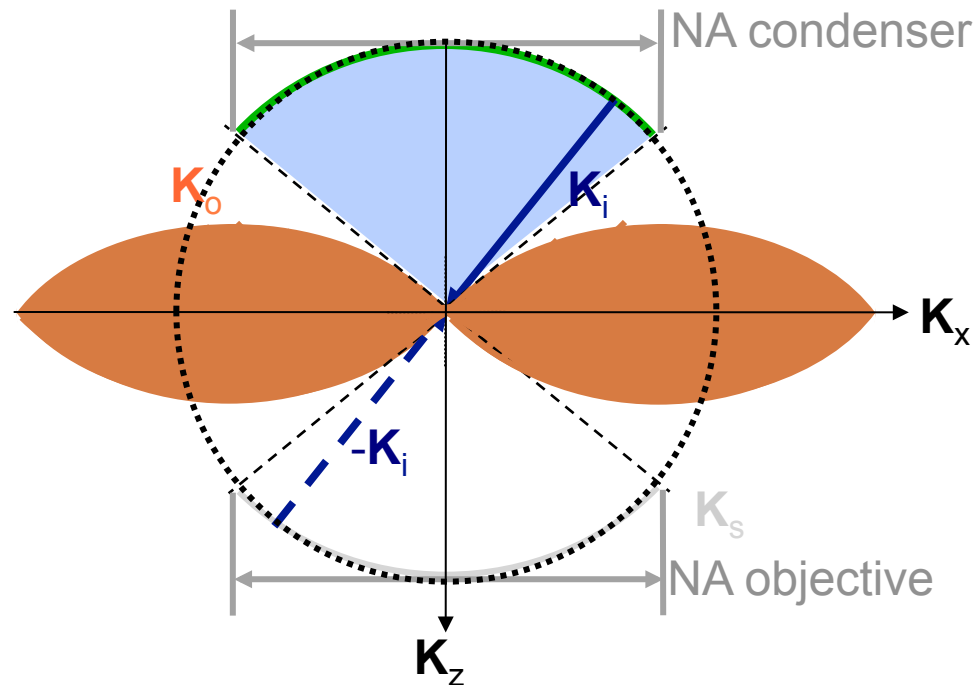
Bon *et al.*, Enhanced 3D spatial resolution in quantitative phase microscopy using spatially incoherent illumination, **Opt. Express**, 22(7), 2014

- Confocal illumination & structured illumination / structured refractive index modulation

Chowdhury & Izzat, Structured illumination quantitative phase microscopy for enhanced resolution amplitude and phase imaging, **Biomed. Opt. Express**, 4(10), 2013

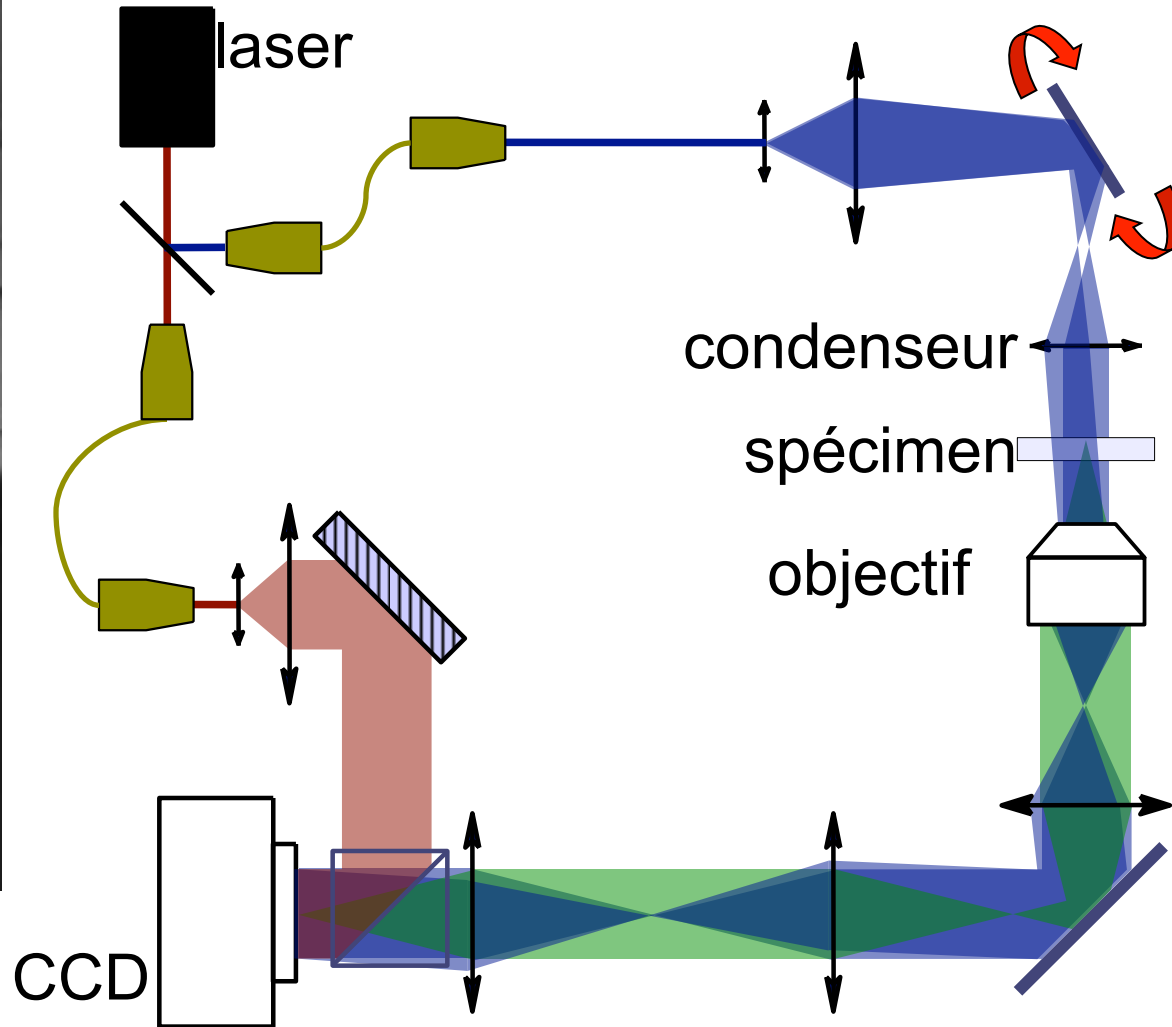
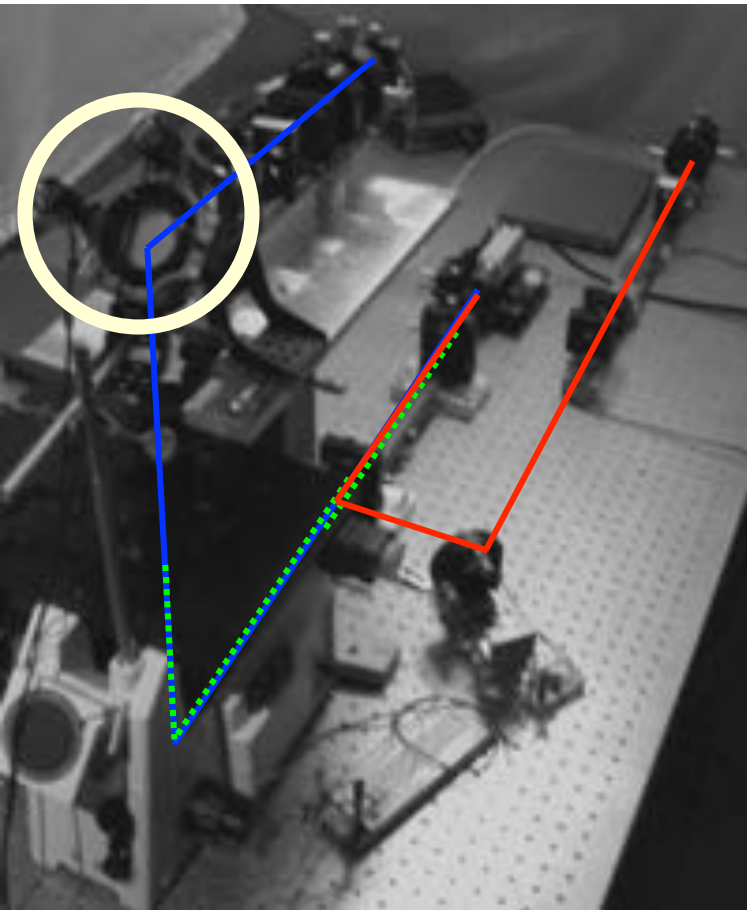
Liu *et al.*, Quantitative phase-contrast confocal microscope, **Opt. Express**, 22(15), 2014

Tomography by Illumination Rotation



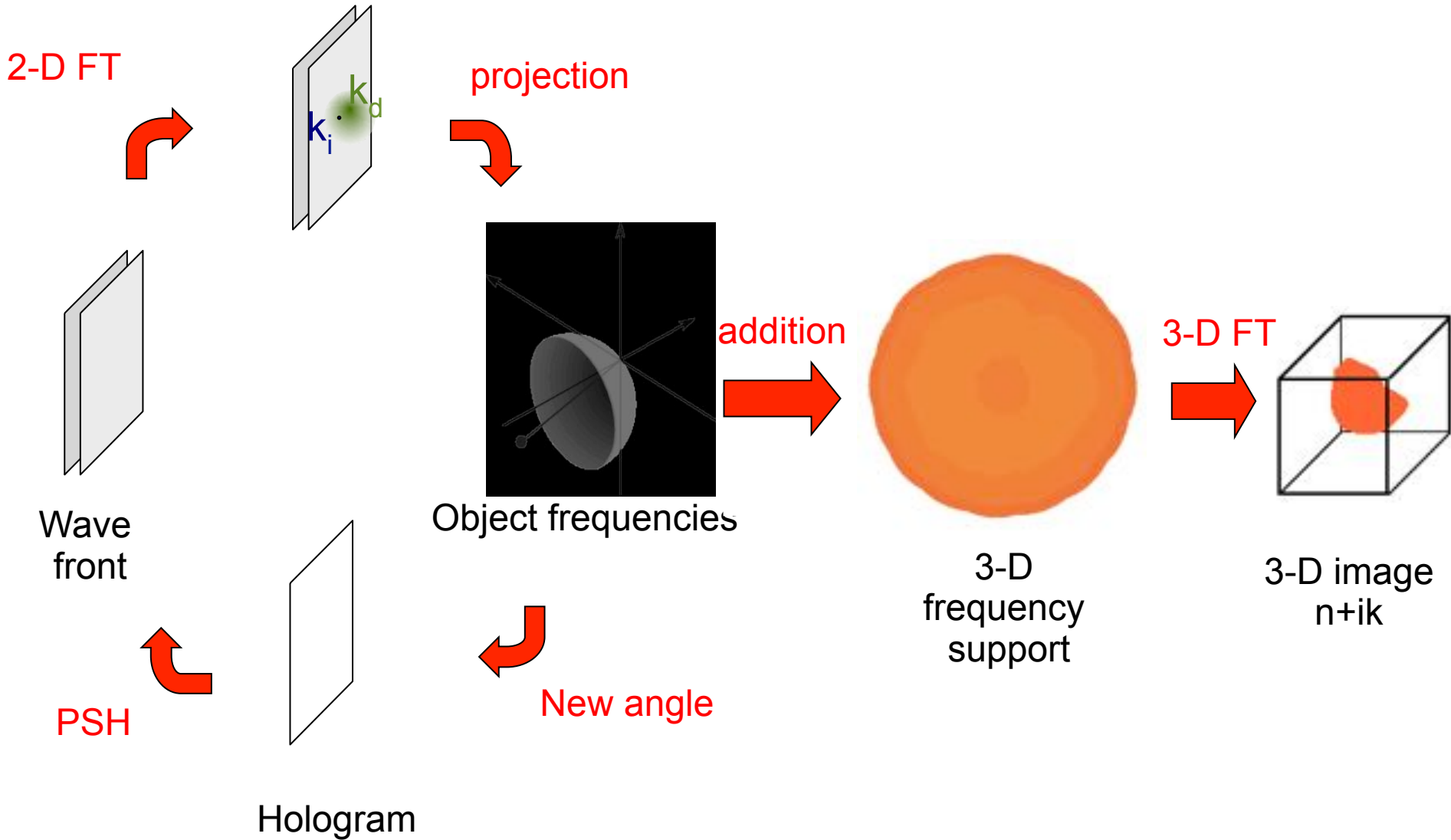
- Different angles of illumination \Rightarrow other object frequencies
- Large number of angles
 \Rightarrow Extended and filled frequency support
- **Objective numerical aperture \Rightarrow Limitation of the detection angle**
- **Condenser numerical aperture \Rightarrow Limitation of the illumination angle**

Tomographic Microscopy: Transmission



Holographic microscopy and diffractive microtomography of transparent samples, M. Debailleul, *et al.*, *Meas. Sci. Technol.* **19**, 074009 (2008)

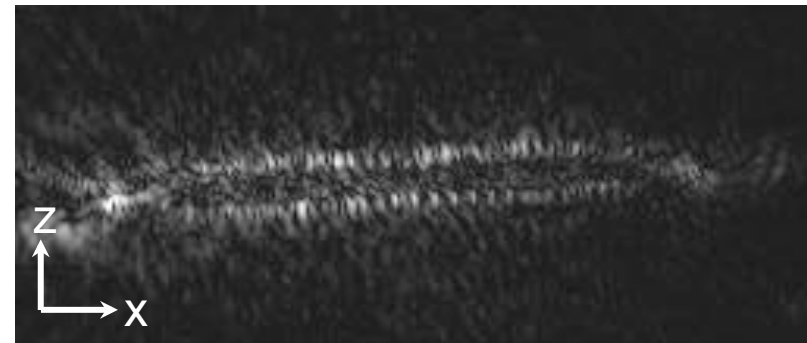
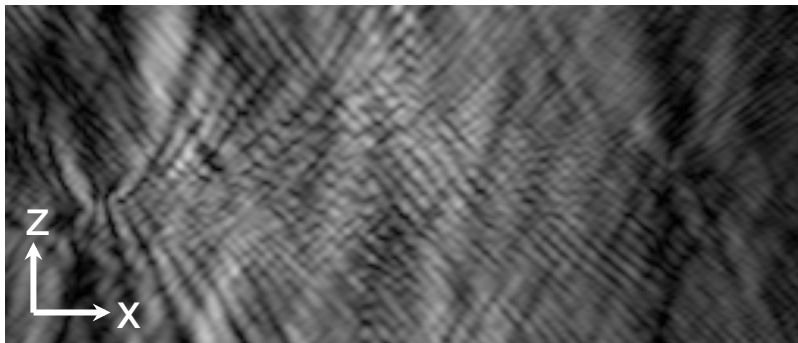
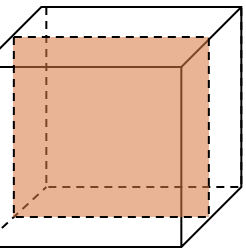
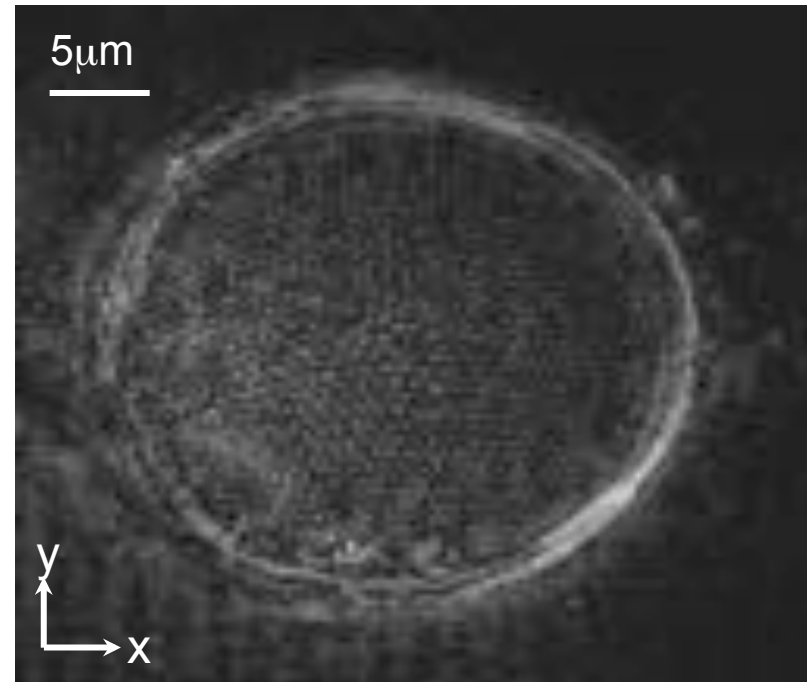
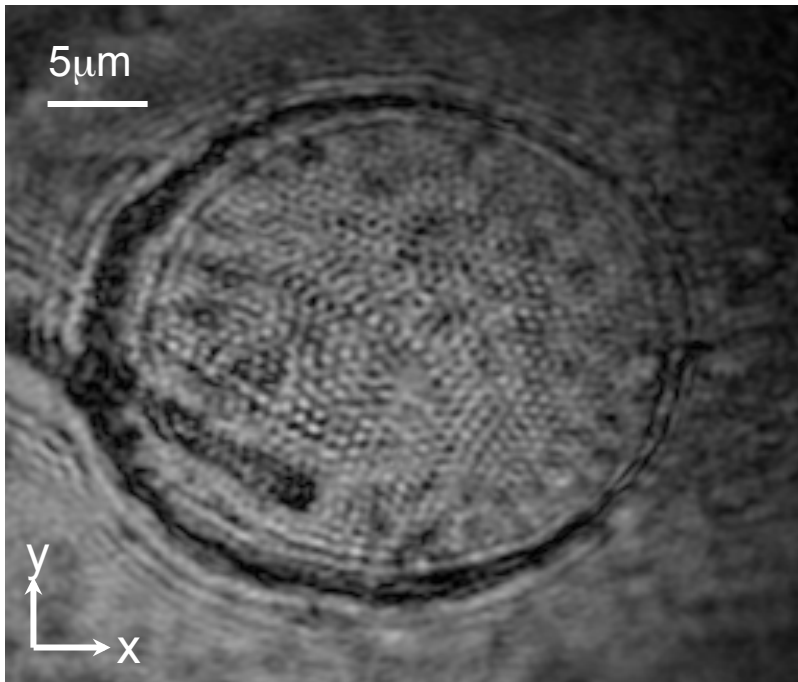
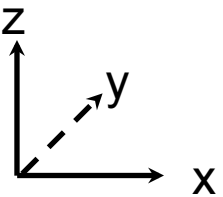
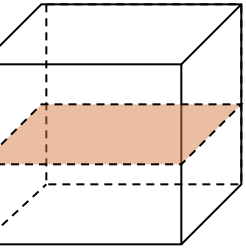
Object Reconstruction



Holography / Tomography

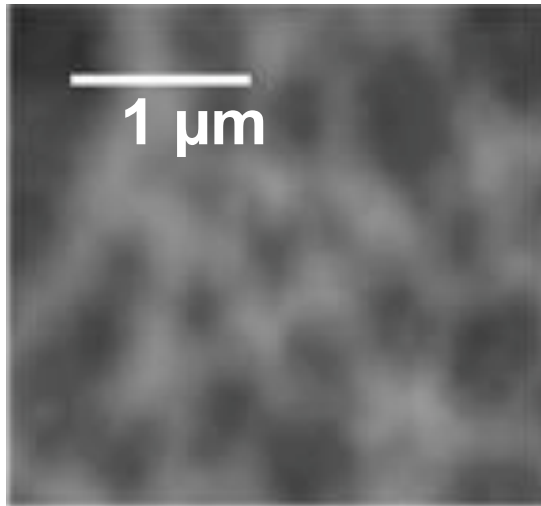
1 angle

372 angles

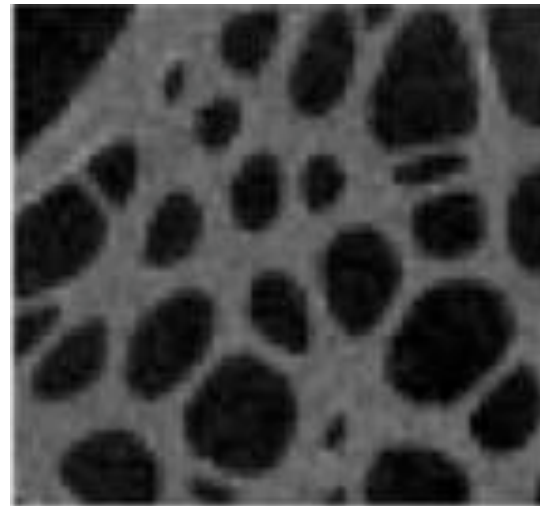


Carbon Mesh

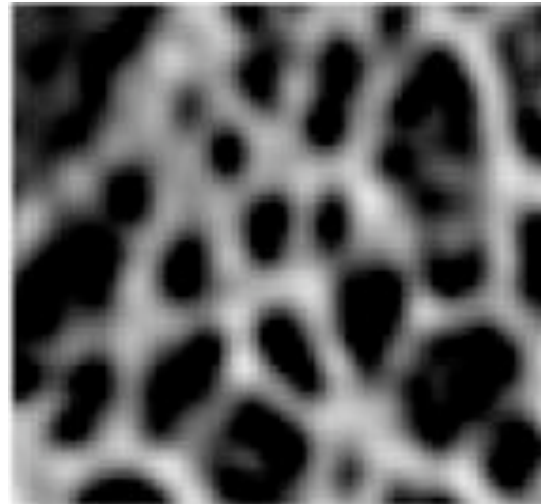
WF



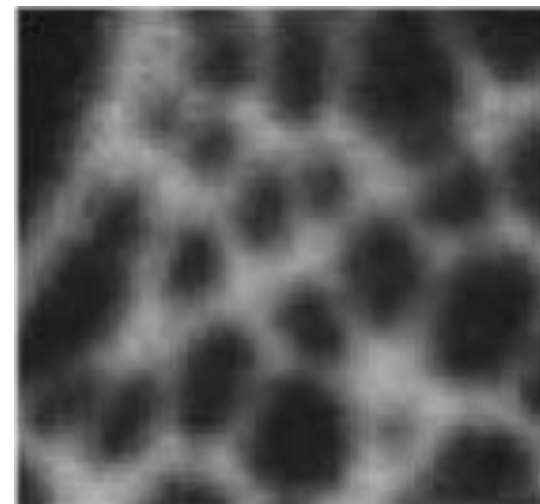
SEM



TDM



LSCM



$R_{\text{exp}}=130\text{nm}$
 $R_{\text{the}}=113\text{nm}$
 $\lambda_{\text{det}}=633\text{nm}$

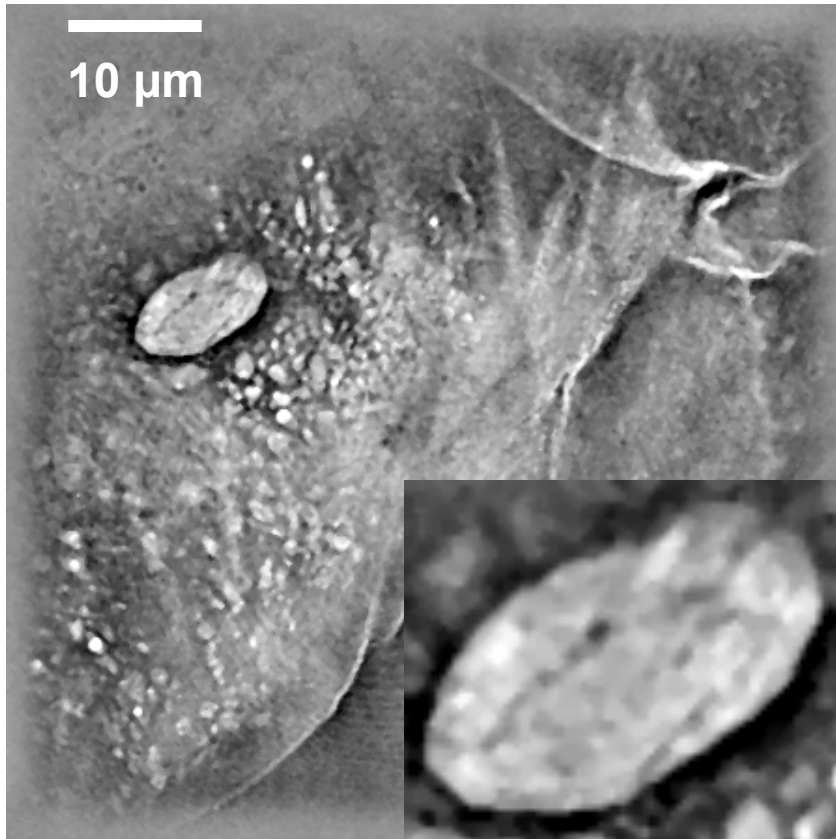
$R_{\text{exp}}=200\text{nm}$
 $\lambda_{\text{exc}}=543\text{nm}$
 $\lambda_{\text{det}}>560\text{nm}$

High-resolution three-dimensional tomographic diffractive microscopy
of transparent inorganic and biological samples

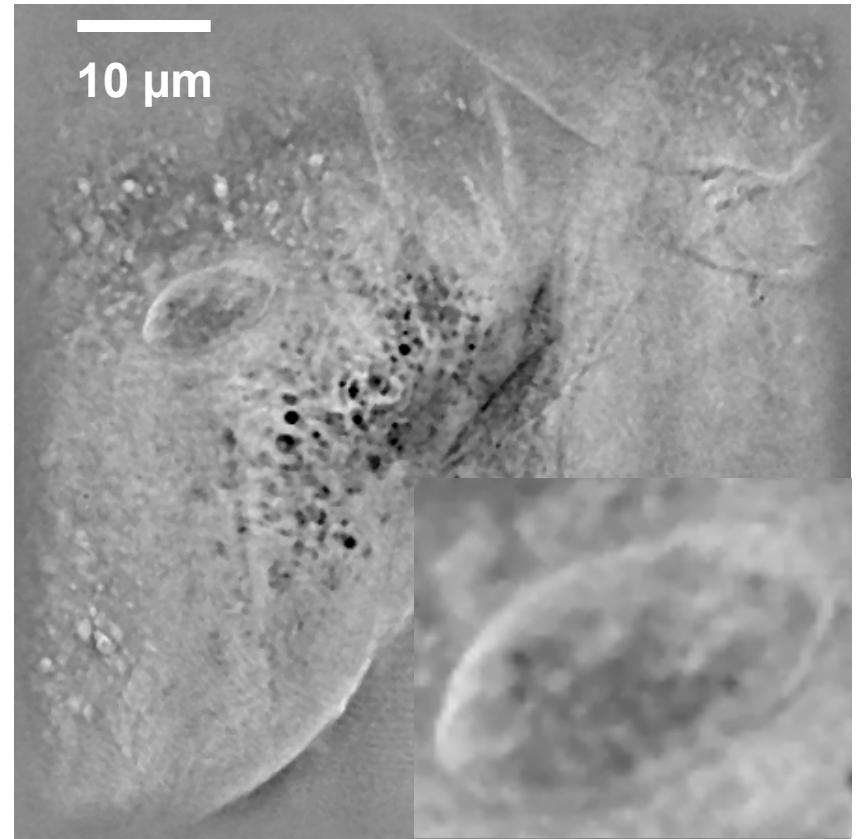
M. Debailleul, *et al.*, *Opt. Lett.* **34**, p. 79 (2009)

Tomography => Index of Refraction

Epithelial cells



**Indice
Real part**



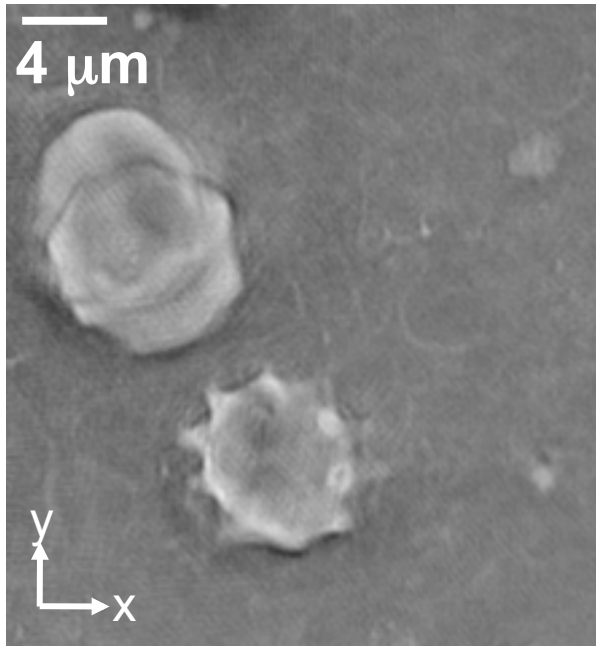
**Indice
Imaginary part**

High resolution tomographic diffractive microscopy of biological samples

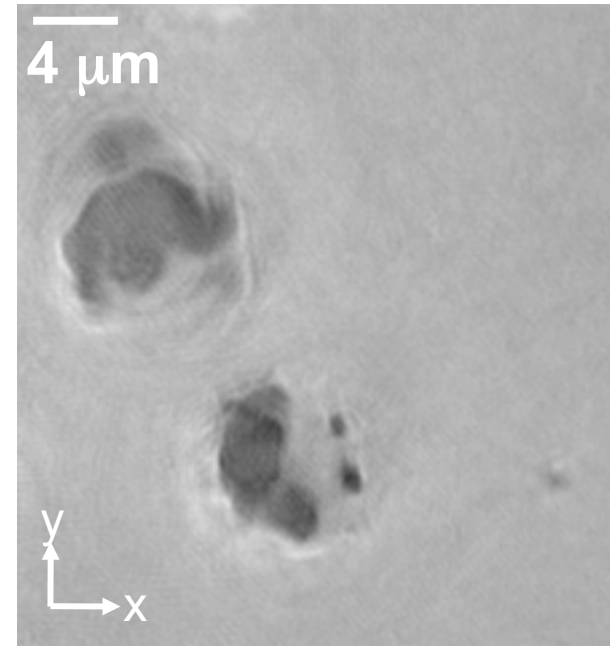
M. Sarmis, *et al.*, J. Biophotonics 3, p. 462 (2010)

Tomography => Index of Refraction

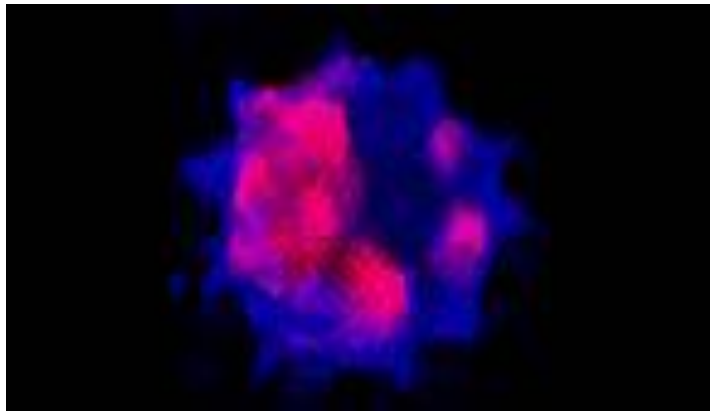
Granulocytes



Refraction

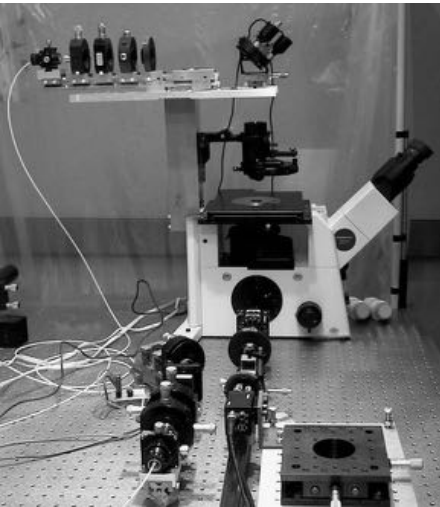


Absorption

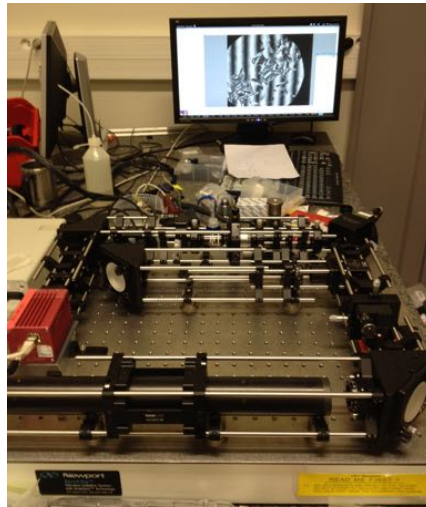


False colour rendering:
Red : absorption
Blue : refraction

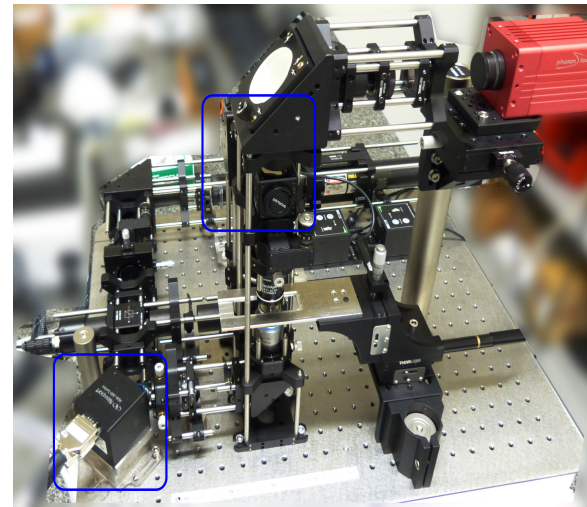
Solutions de labo



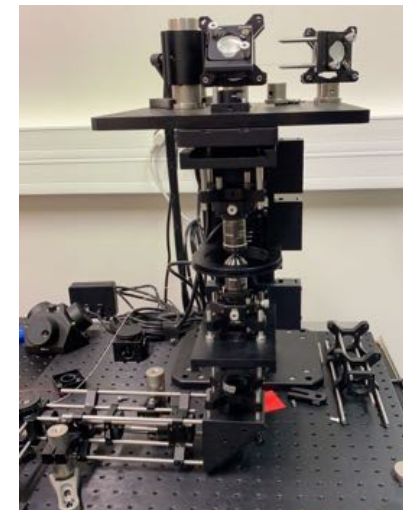
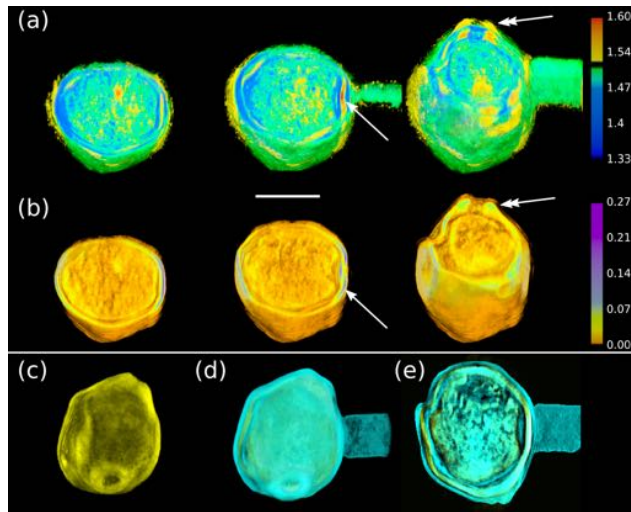
First prototype



Fast Compact Setup
MiFoBio 2014-2016



Compact Setup MiFoBio 2012



New System

Commercially available!



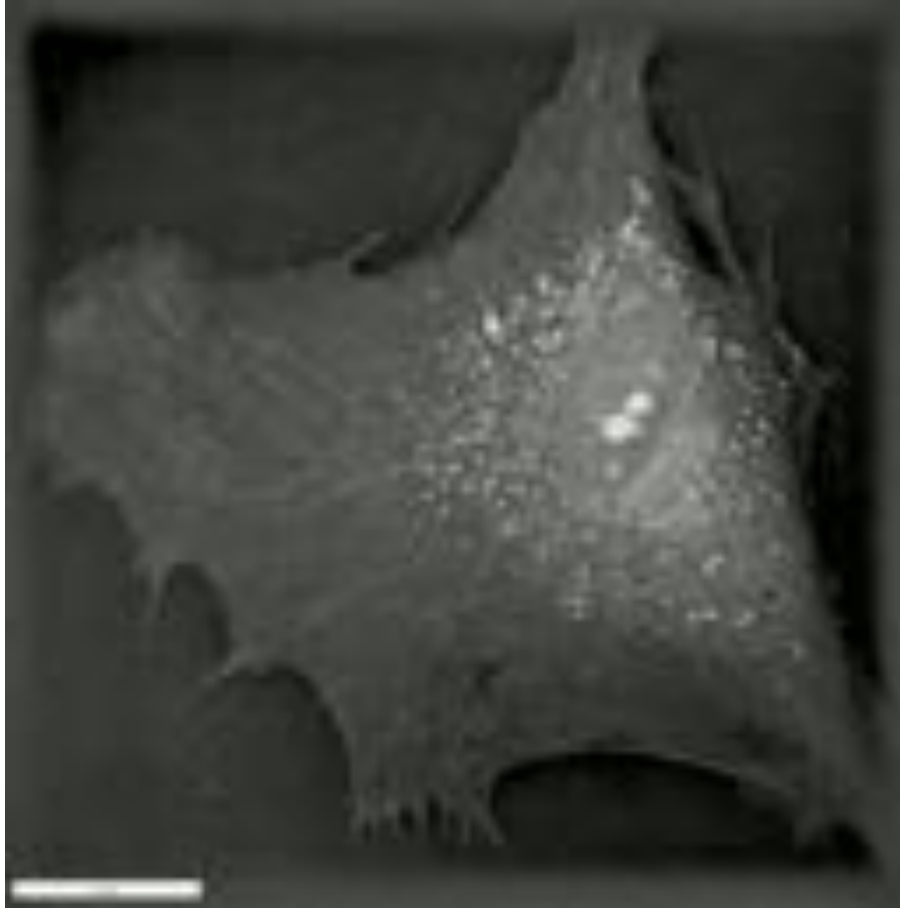
NANOLIVE 
Looking inside life



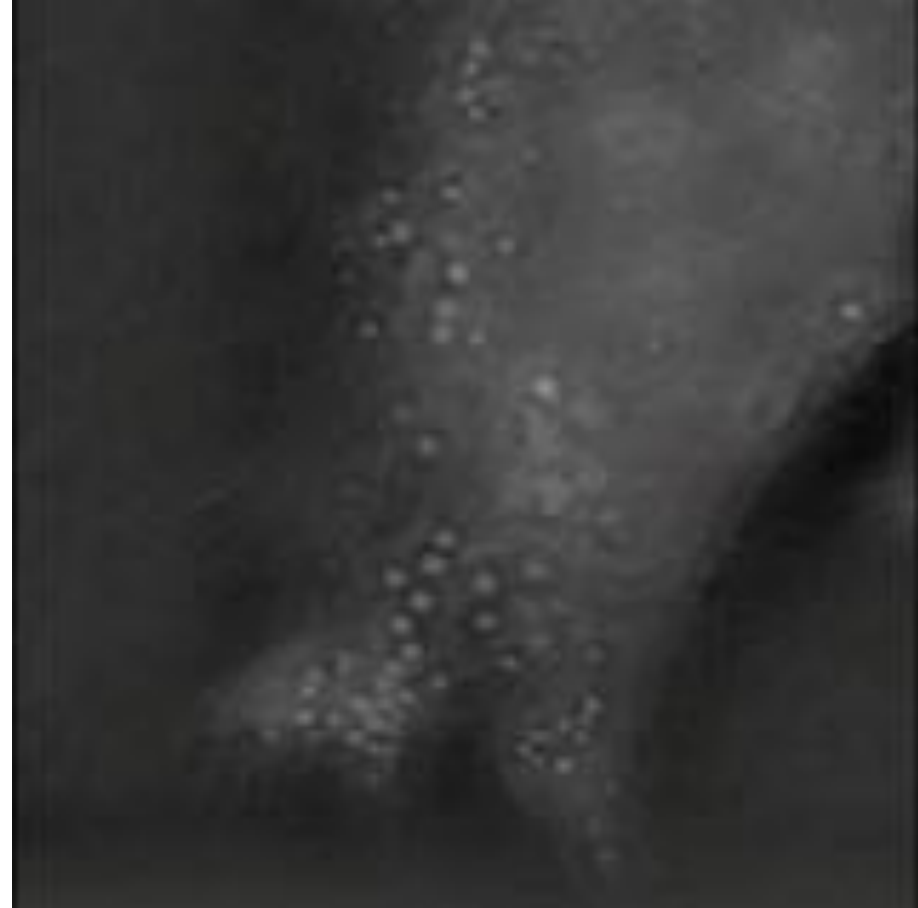
 **TOMOCUBE**

**See their website for interesting applications
Several active groups in the world (Korea, Poland, Taiwan,
France, Germany, Italy...).**

Applications <http://nanolive.ch>



cellular morphological changes
induced by drug treatment

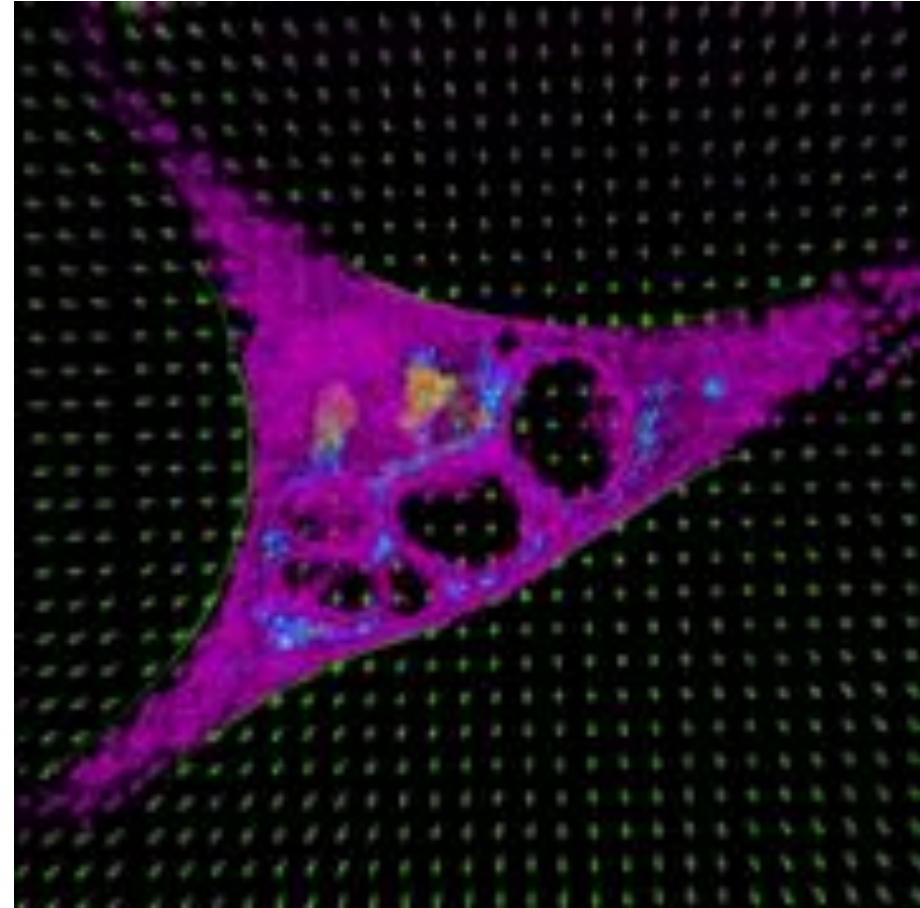


nanodiamonds internalization
& 3D distribution in living cells

Applications <http://nanolive.ch>



Fission yeast
(*Schizosaccharomyces pombe*)
during division



Fibroblast reticular cell seeded
on glass nanopillars

Applications <http://nanolive.ch>

Oncology &
Immuno-Oncology

Mitochondria & Cell
Metabolism

Stem cells

Cytotoxicity

Single cell
characterization

Immunology

Neuroscience

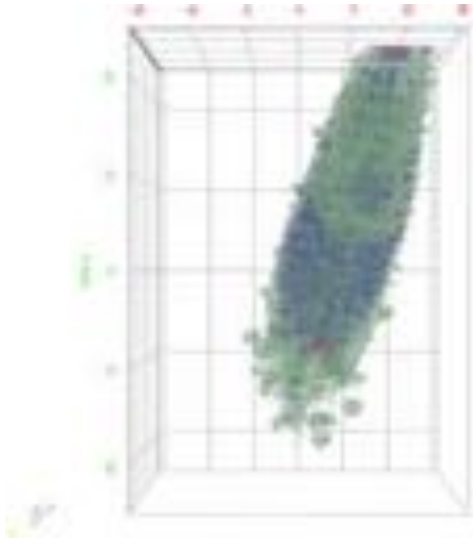
Microbiology

Cell-material
interactions

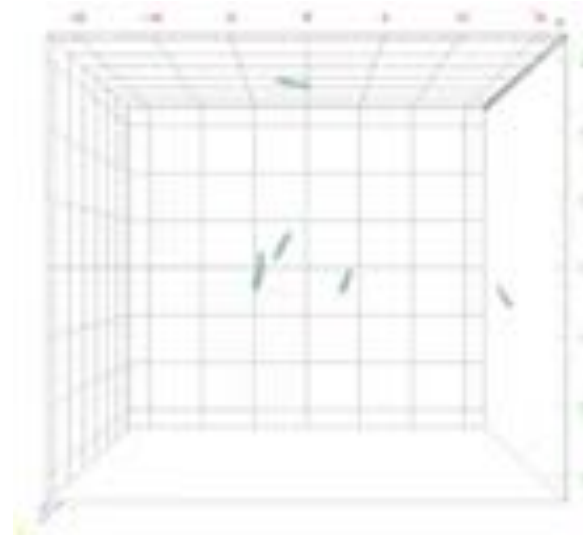
Model Organisms

Applications <http://www.tomocube.com>

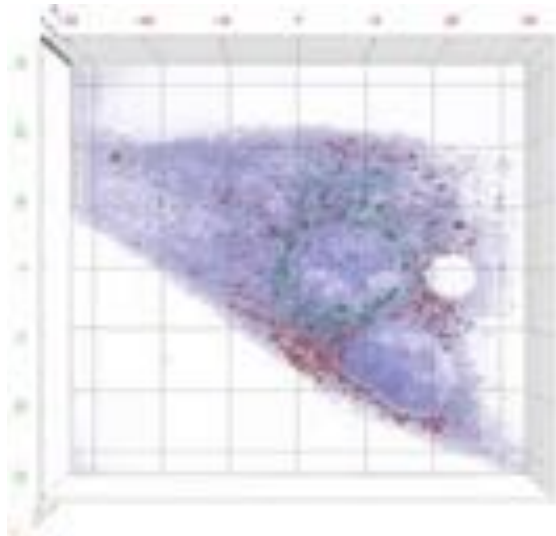
Cell
apoptosis



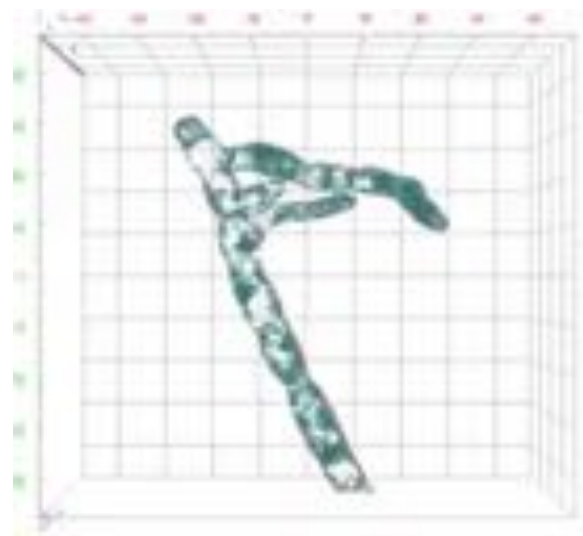
Bacterial
growth



HeLa cell



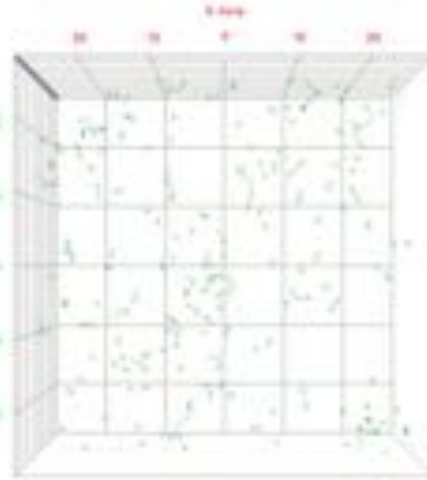
Microalgae



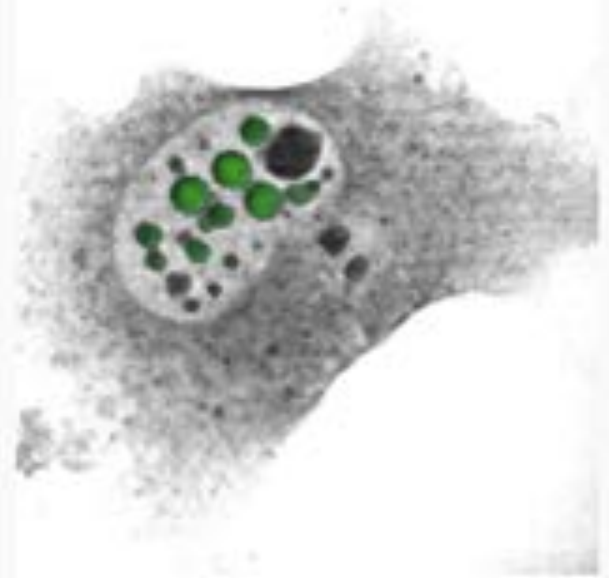
Applications <http://www.tomocube.com>



Lipid Quantification

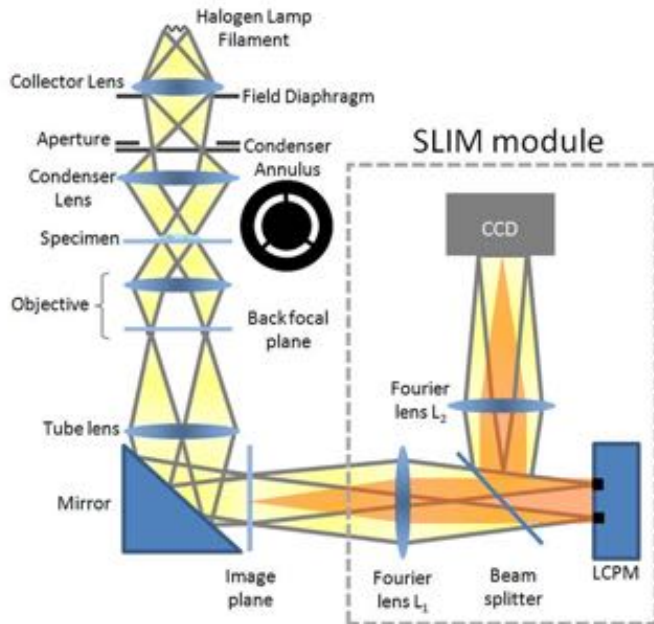


Bacteria Analysis



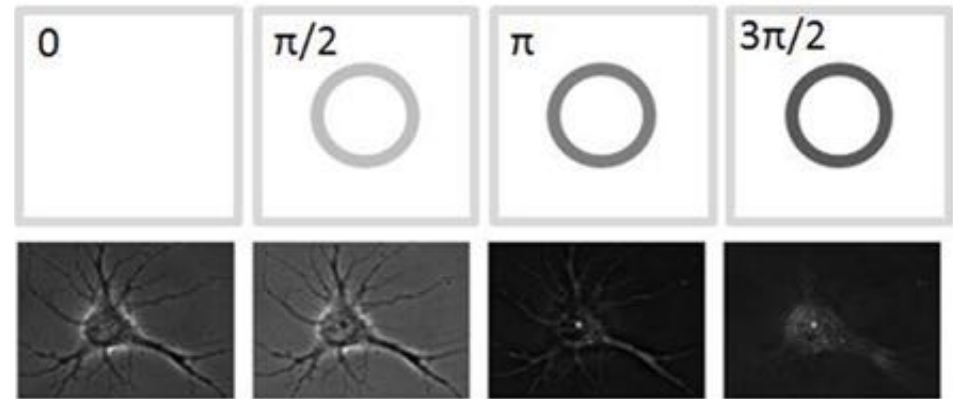
Phase Separation

White-light diffraction tomography

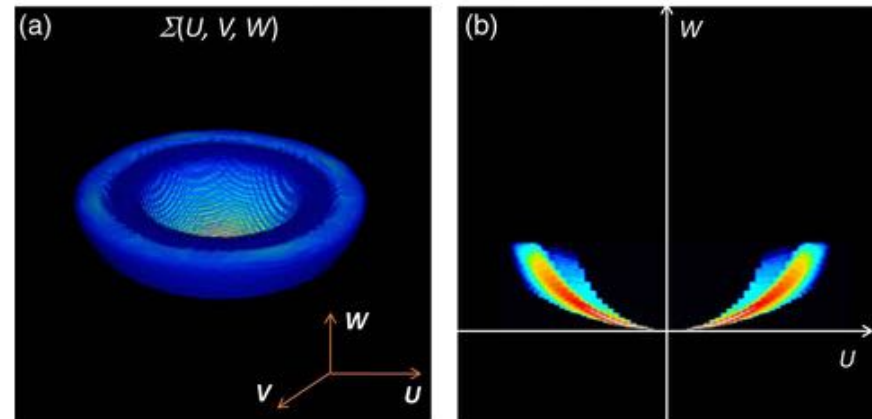


+ Z-scanning
+ data processing

Phase rings



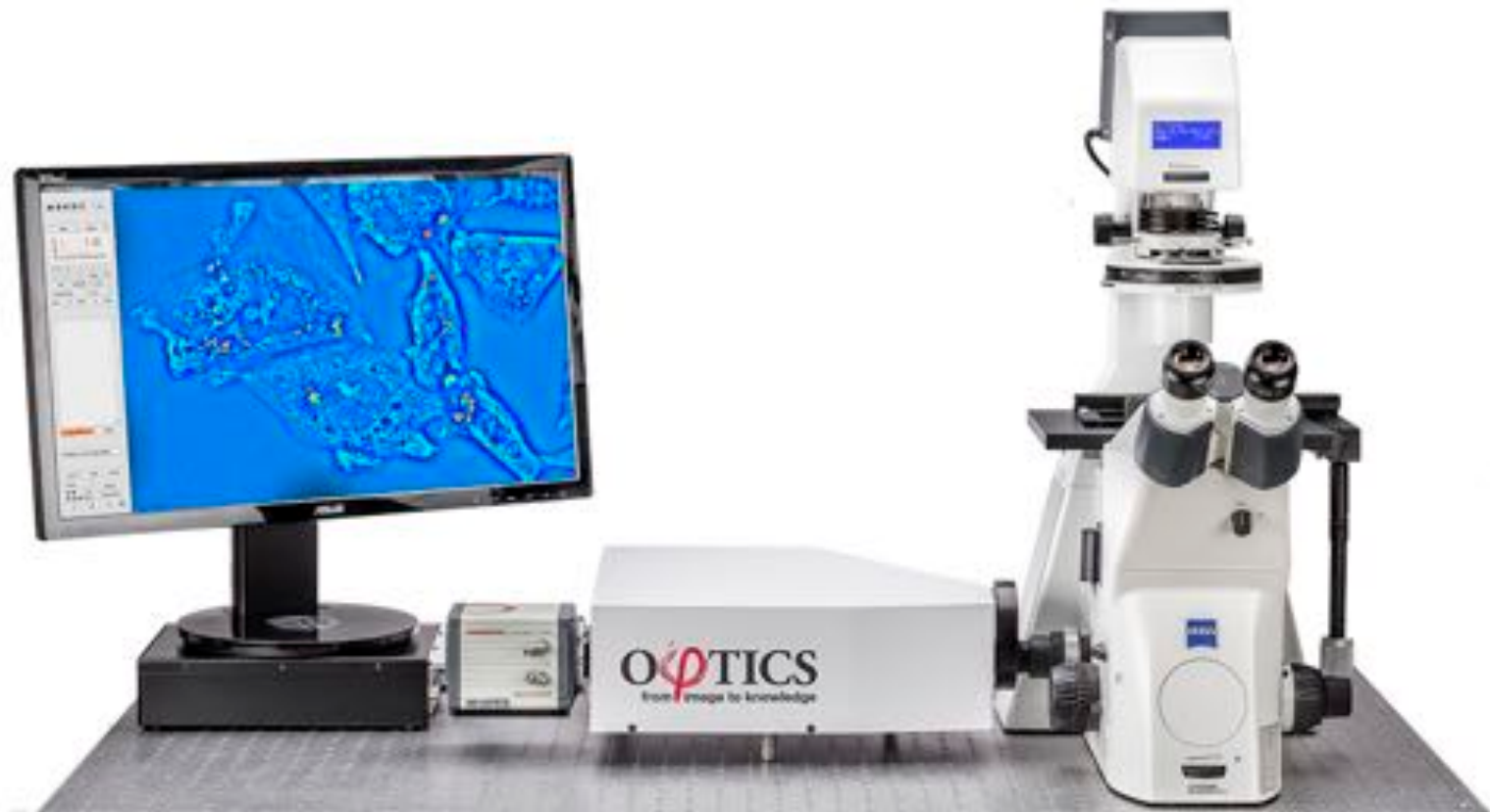
Coherent transfer function



White-light diffraction tomography of unlabelled live cells

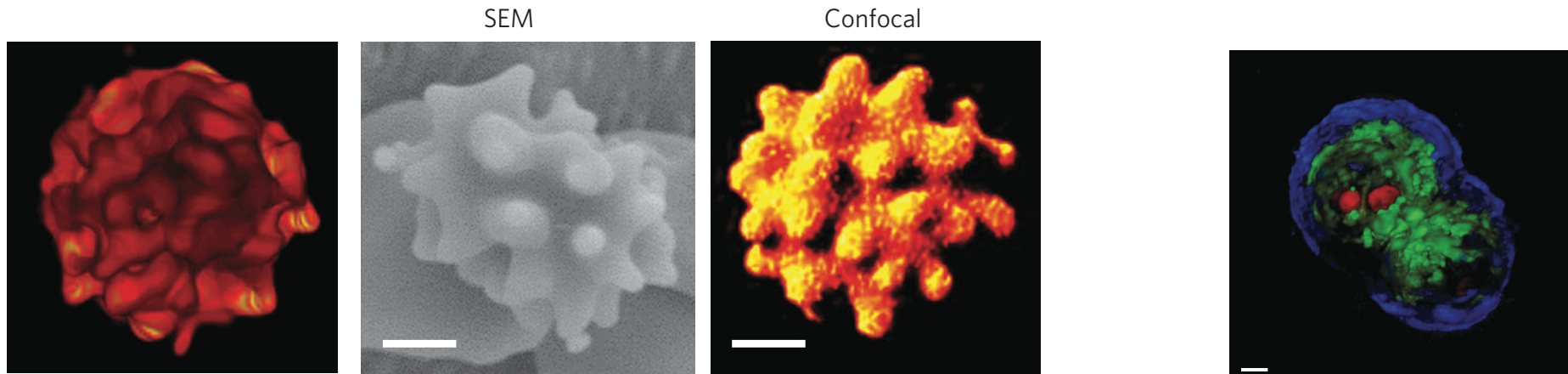
T. Kim, *et al.* Nature Photonics 8, p. 256 (2014)

Commercially available!



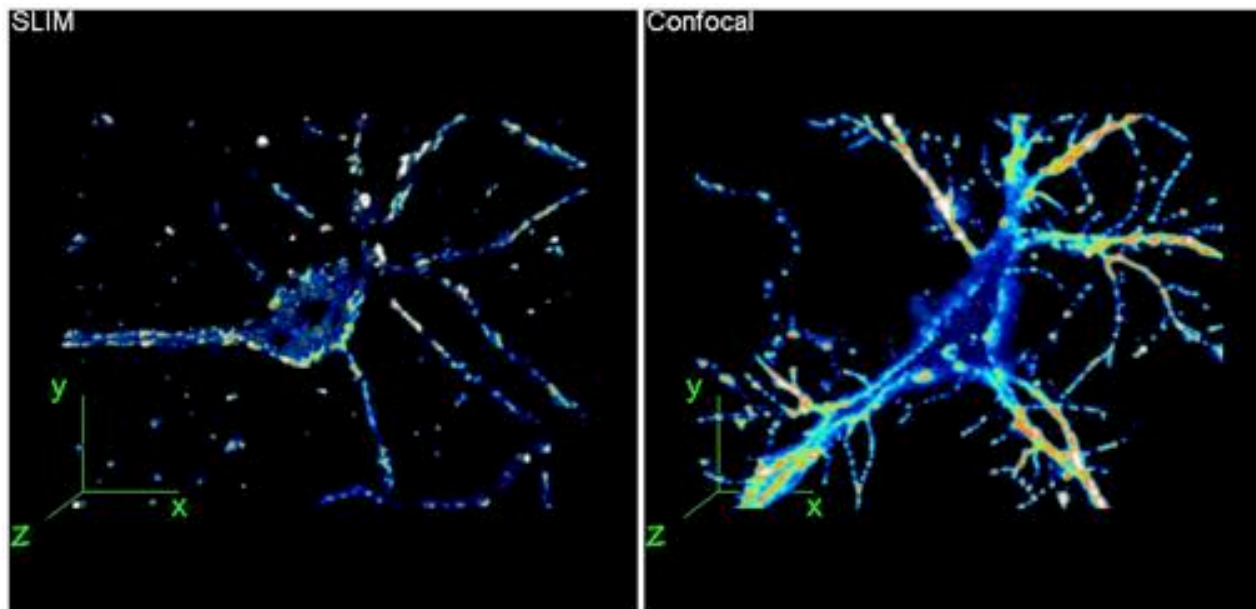
See their website for interesting applications

Applications <http://phioptics.com>



Spiculated RBC

HT29 cell



Live neuron

Applications <http://phioptics.com>

3D Tomography

Home / Applications / 3D Tomography

3D Tomography

Cell Dynamics

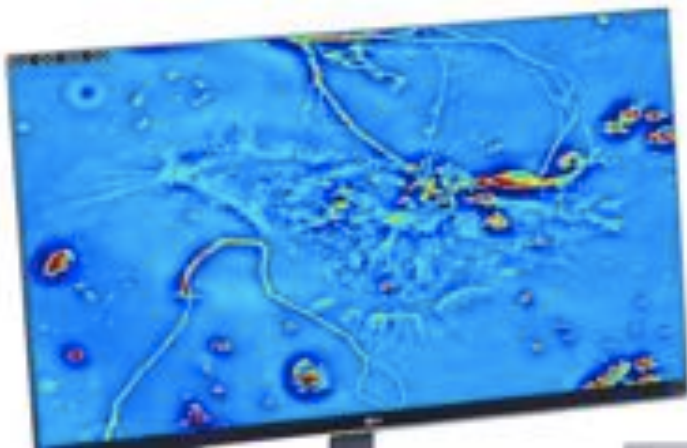
Cell Growth

Neuroscience

Particle Analysis

Tissue Imaging

3D cell tomography with SLIM



Limits of Label-free VS fluorescence

Sensitivity

→ Not a single molecule approach...

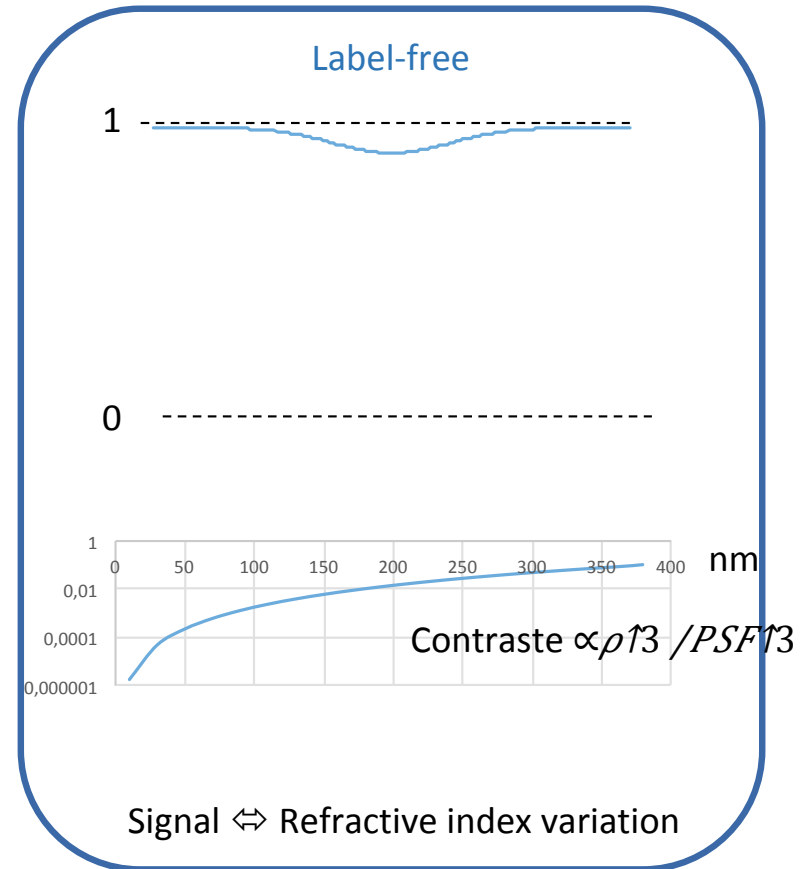
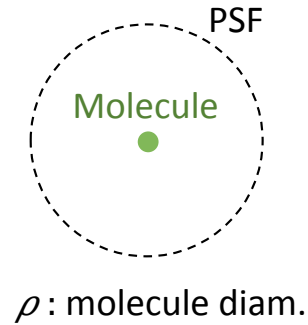
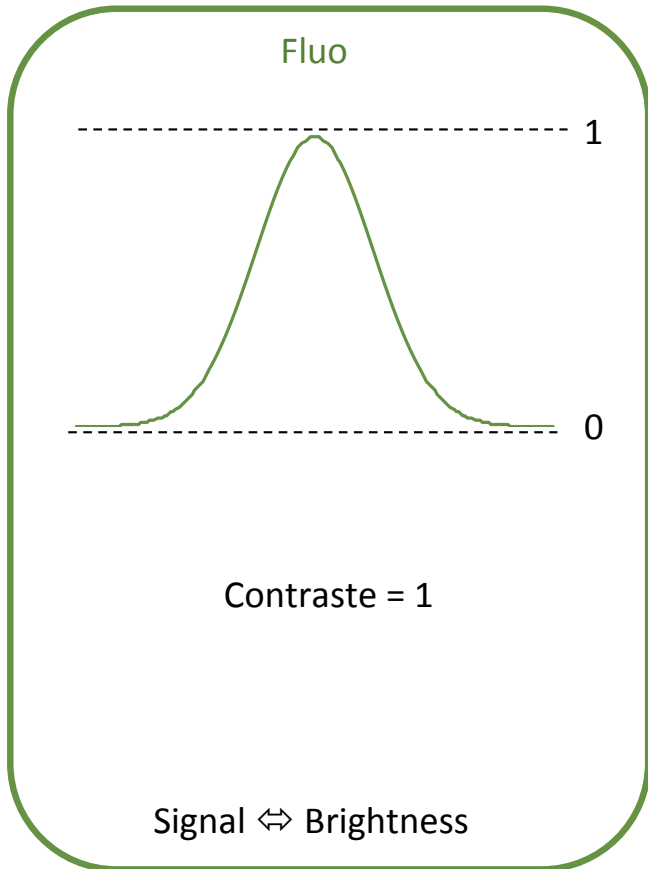
Resolution limit

→ Initial resolution = 50% of fluo. resolution

Selectivity/Specificity

→ How to identify and contrast a molecular type

Sensitivity



Sensitivity enhancement ?

- Signal = shot-noise limited → more photons! → alternative sensor

Hosseini *et al.*, Pushing phase and amplitude sensitivity limits in interferometric microscopy, **Optics Letters**, 41(7), 2016

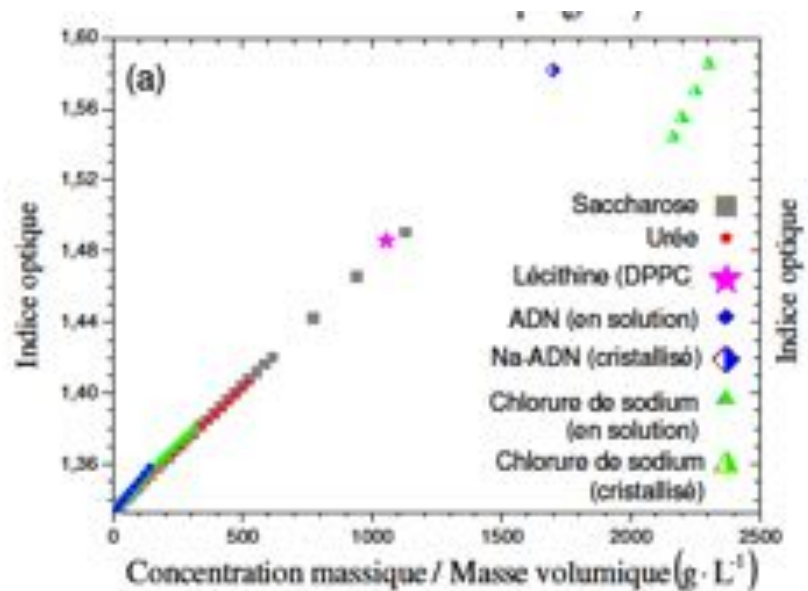
Roose-Amsaleg *et al.*, Utilization of interferometric light microscopy for the rapid analysis of virus abundance in a river, **Research in microbiology**, 168(5), 2017

- Dark field

Martinez-Marrades *et al.*, Stochastic 3D optical mapping by holographic localization of Brownian scatterers, **Opt. Express**, 22(23), 2014

Selectivity / sensitivity

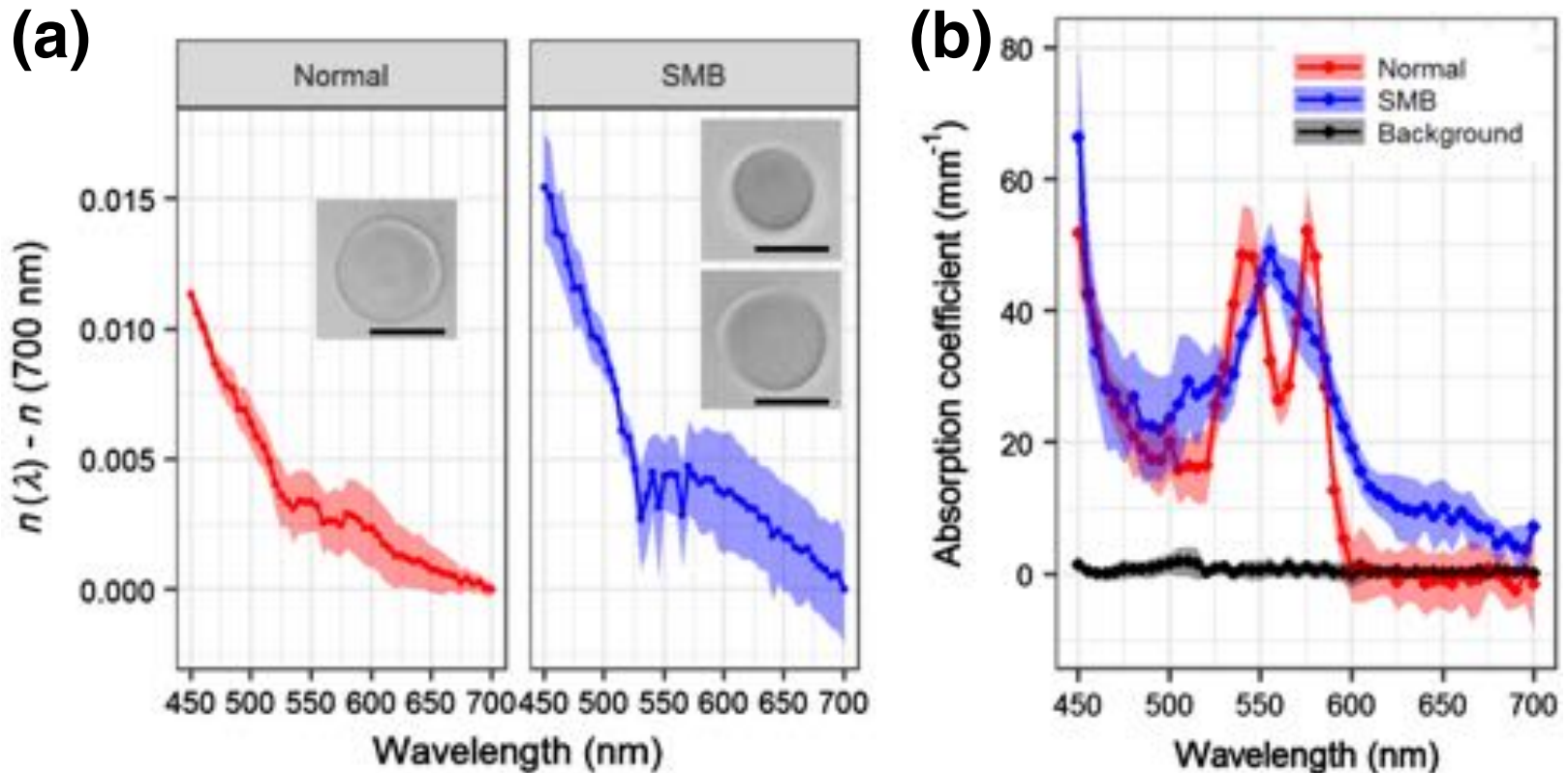
- Signal \Leftrightarrow Refractive index variation ... not specific !



Selectivity / sensitivity enhancement

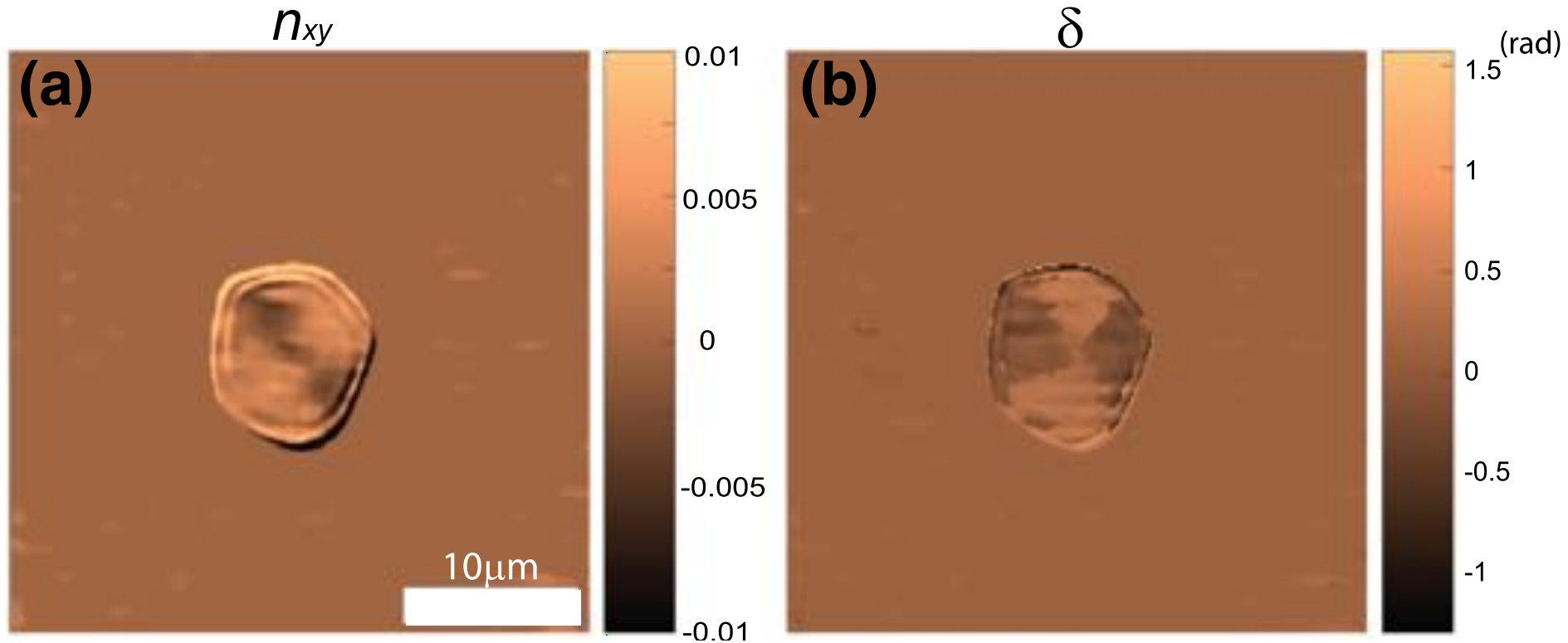
- Polarization → cytoskeleton, collagen...
- High refractive index / absorption / scattering probes (nano-diamond, Gold particle)
- Refractive index specific modulation

Spectroscopic Microtomography



Spectroscopic Microtomography in the Visible Wavelength Range
Y. Sung, Physical Review Applied **10**, 054041 (2018)

Polarized Microtomography



Polarization-sensitive optical diffraction tomography

A. Saba, et al., *Optica* **8**, 402 (2021)

Polarized Microtomography



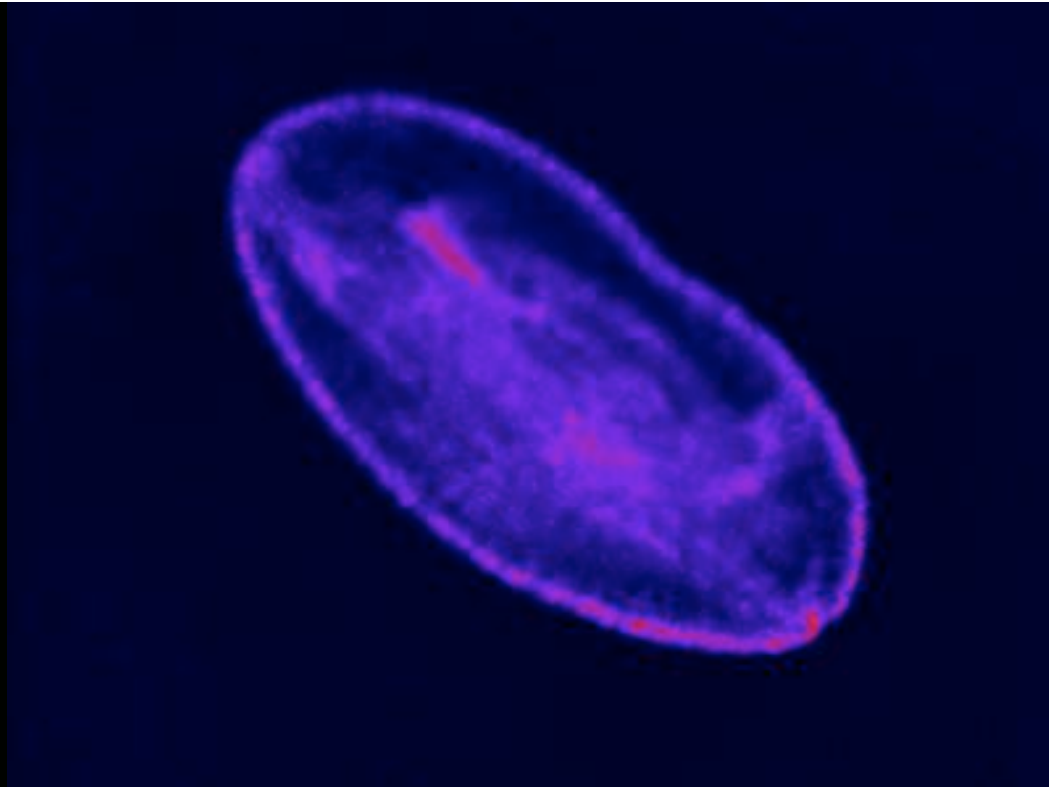
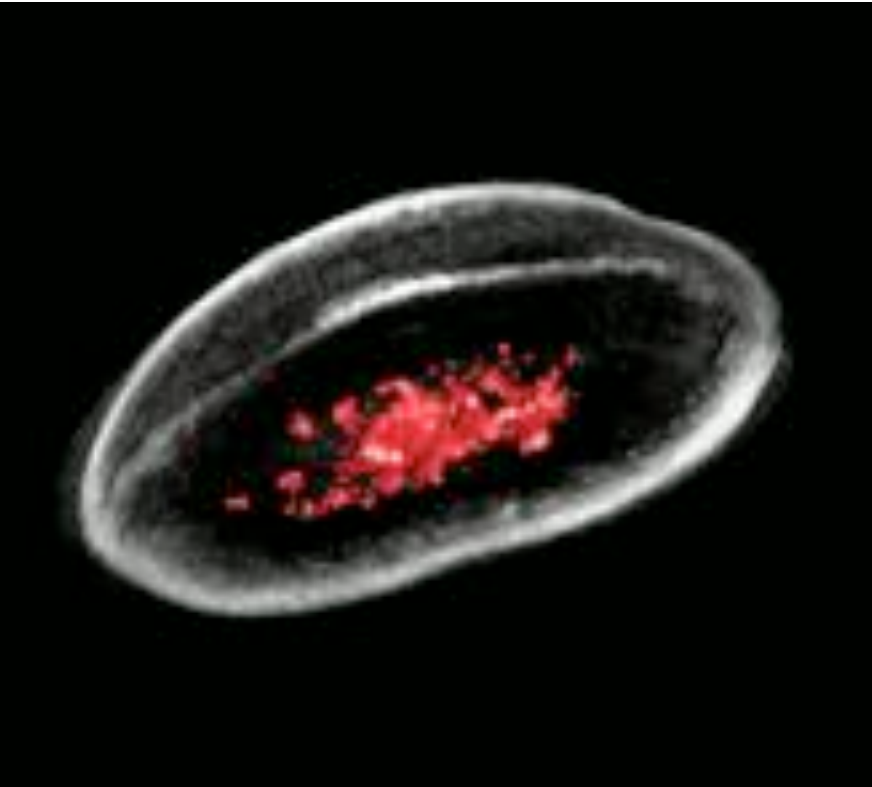
Polarization-sensitive optical diffraction tomography

A. Saba, et al., *Optica* **8**, 402 (2021)

What's next ???

Tomography / Fluorescence Comparison

Snowdrop pollen



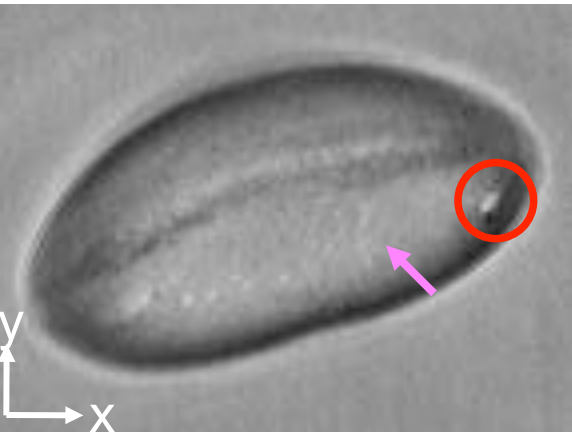
Tomography

red : index $n >$ index immersion medium

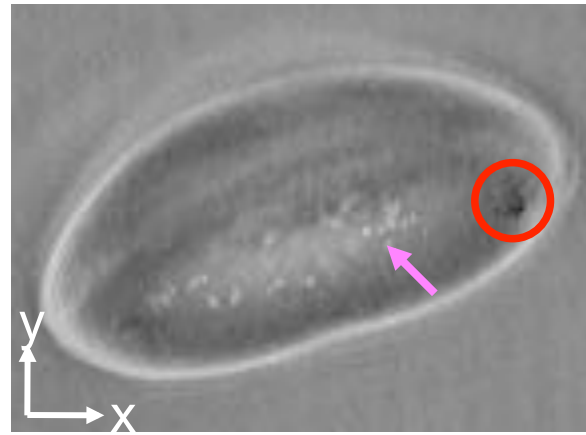
Fluorescence

Non-Isotropic Resolution

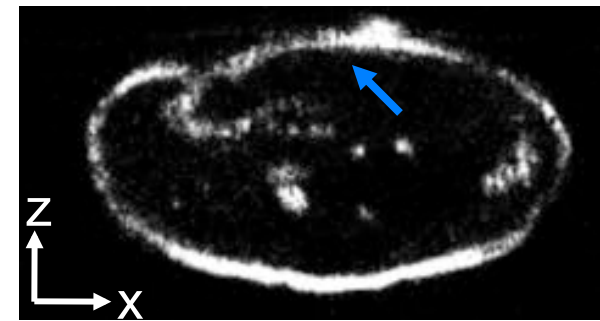
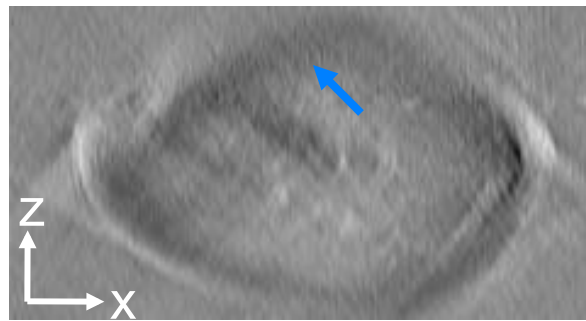
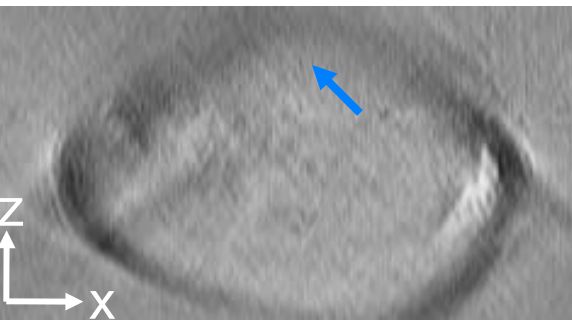
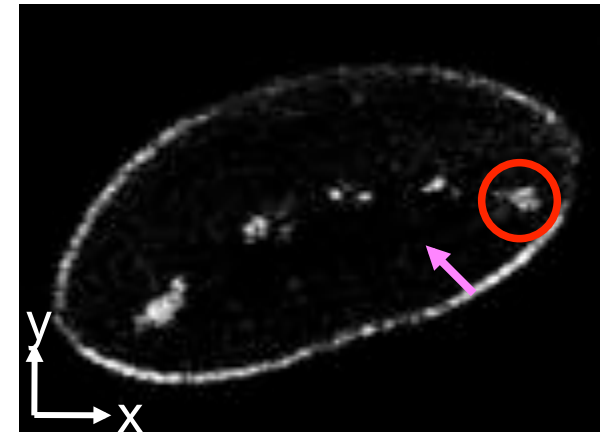
Absorption



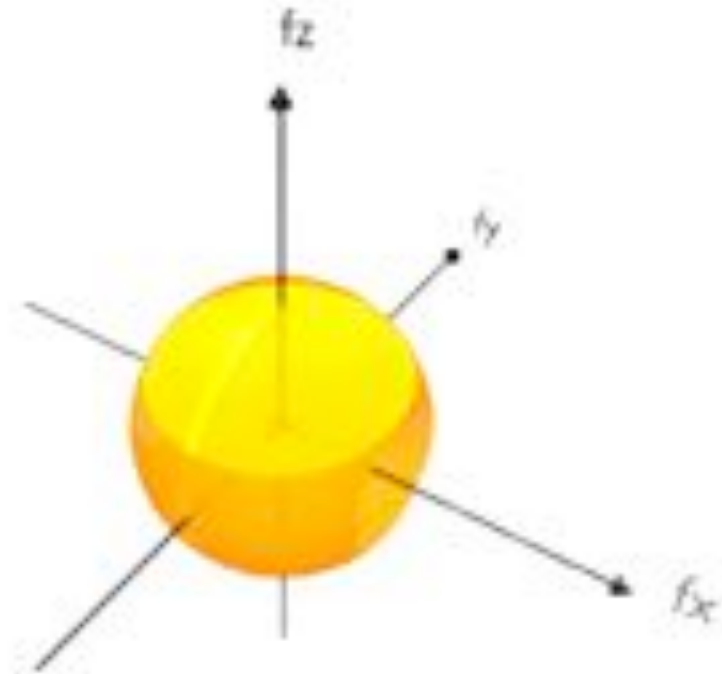
Index of refraction



Autofluorescence



Tomography by Specimen Rotation



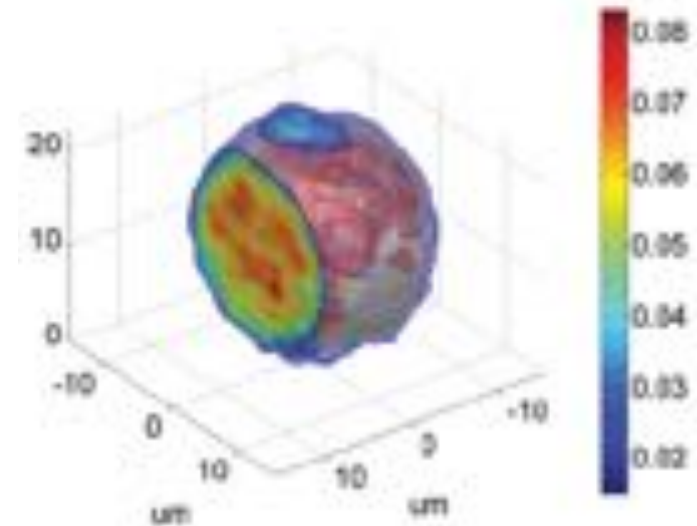
- **Sample rotation may be difficult**
- **low NA \Rightarrow quasi-isotropic, but rather low resolution**

Tomography by Specimen Rotation

Problems and Solutions in 3-D Analysis of Phase Biological Objects by Optical Diffraction Tomography

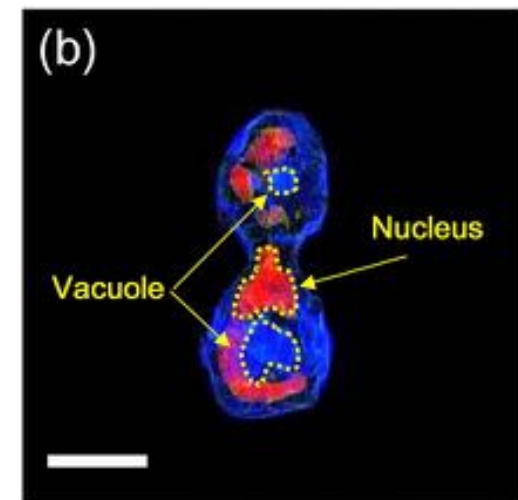
M. Kujawińska, *et al.*,

Int. J. Optomechatronics **8**, p. 357 (2014)



Tomographic phase microscopy with 180° rotation of live cells in suspension by holographic optical tweezers

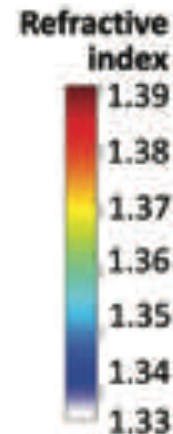
M. Habaza, *et al.*, Opt. Lett. **40**, p. 1881 (2015)



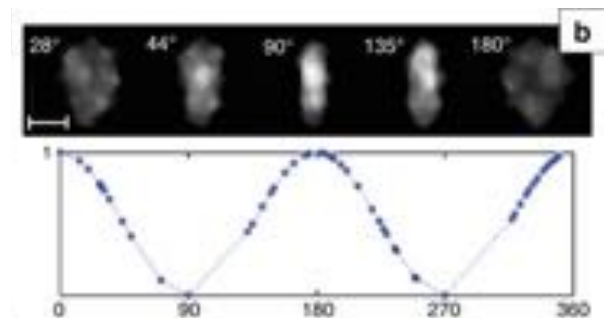
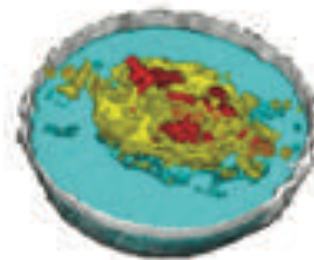
Tomography by Specimen Rotation

Rapid 3D Refractive-Index Imaging of Live Cells in Suspension without Labeling Using Dielectrophoretic Cell Rotation

M. Habaza, *et al.*,
Adv. Sci., paper 1600205 (2016)



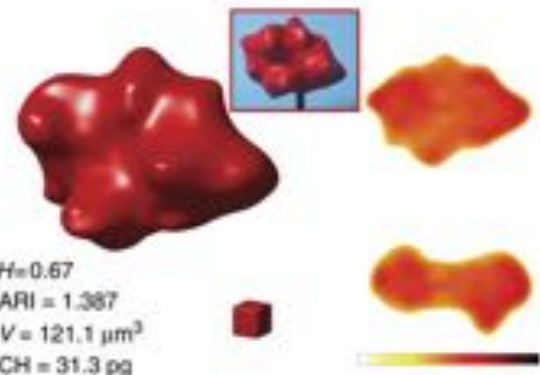
(d)



b

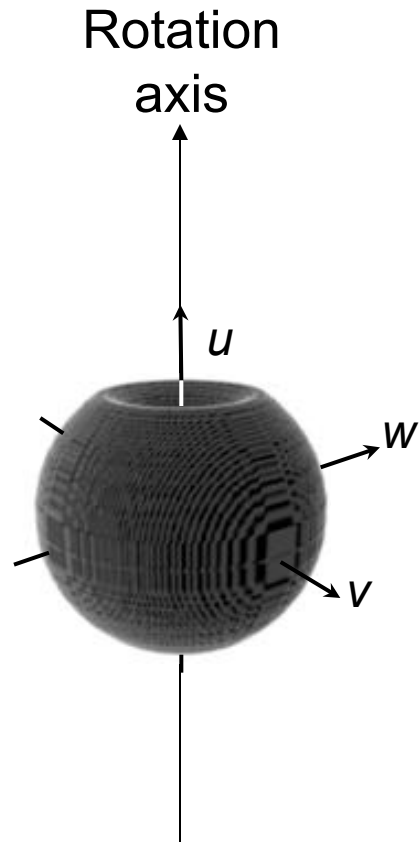
Tomographic flow cytometry by digital holography

F. Merola, *et al.*,
Light: Science & Applications 6,
paper e16241 (2017)



Missing Frequencies

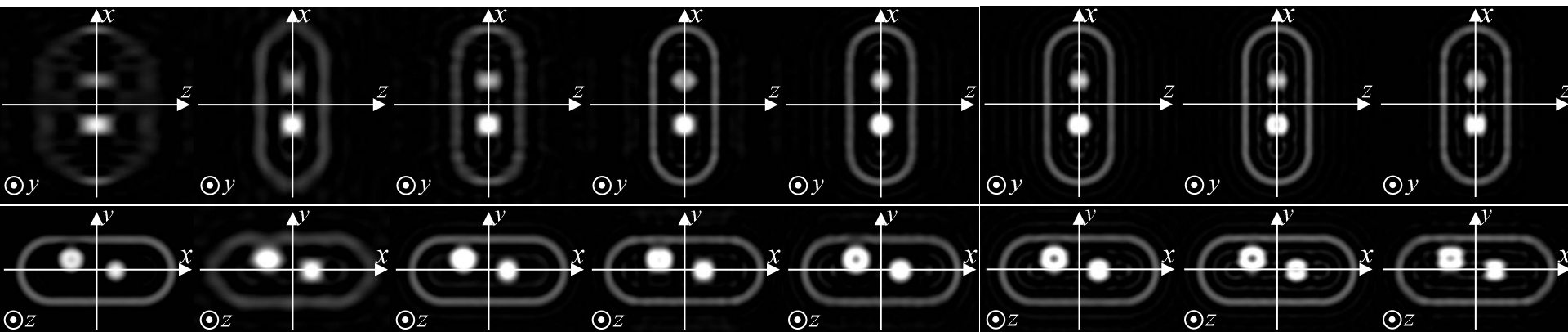
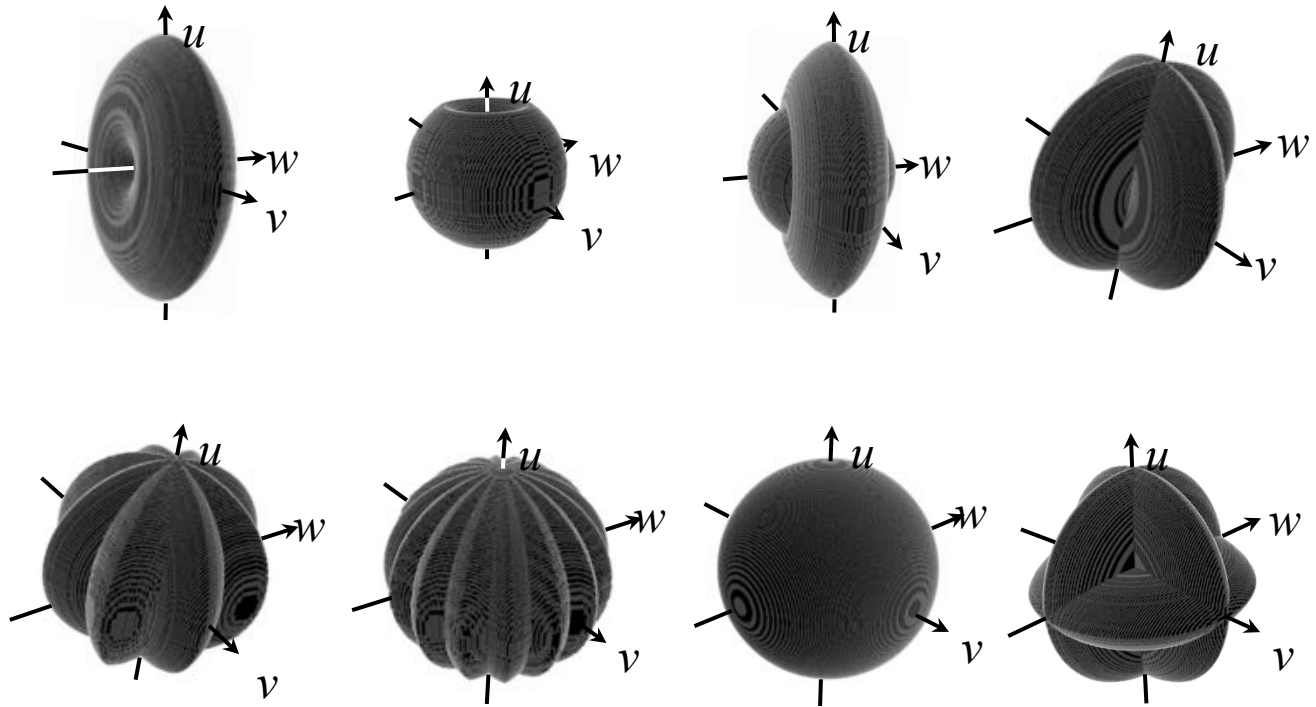
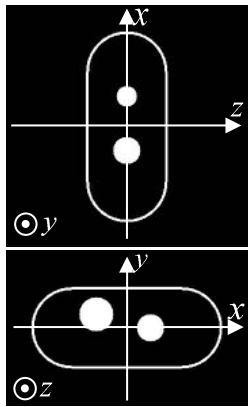
Recorded frequencies Missing part “Missing apple core”



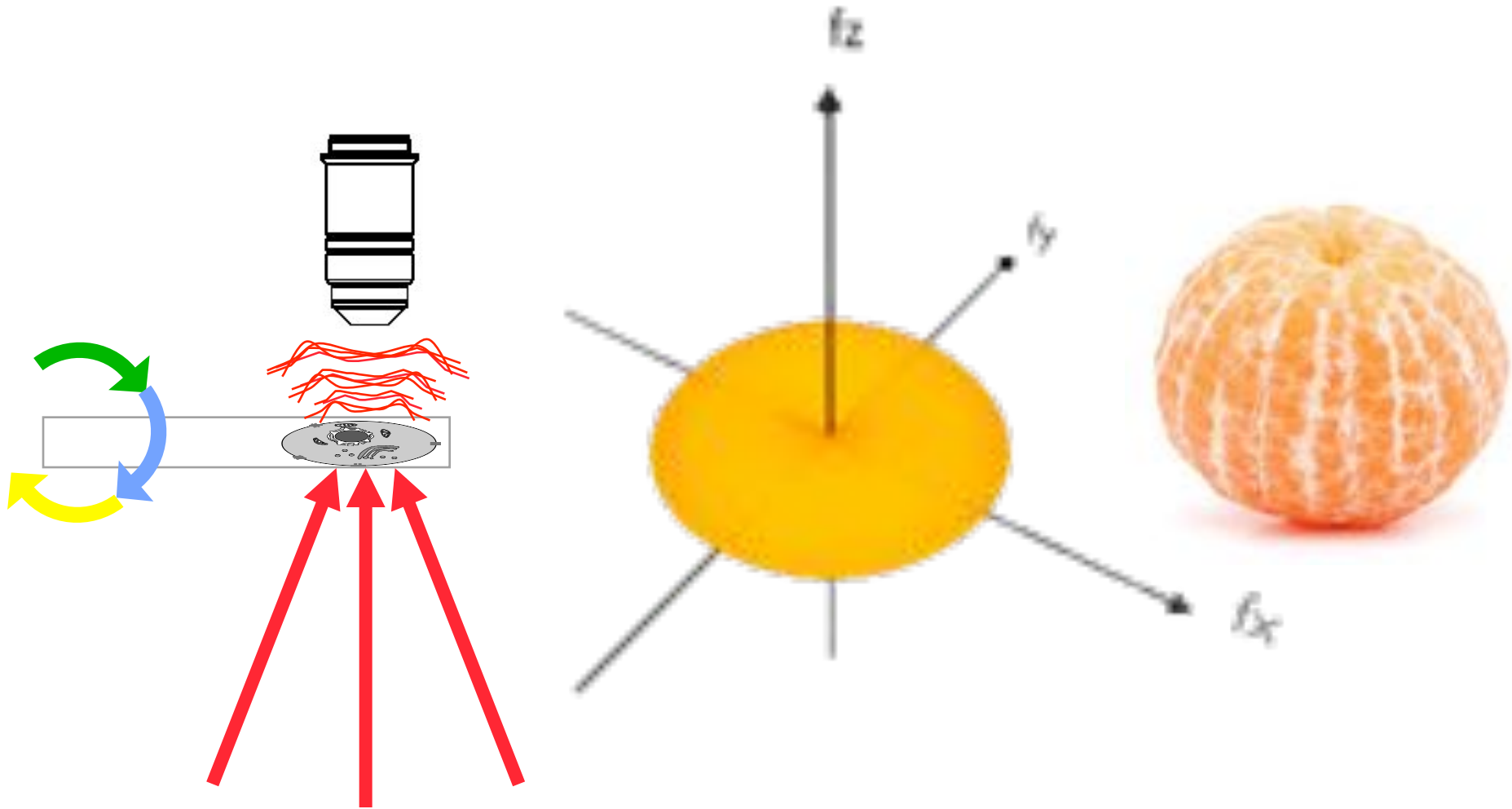
“Diffraction microtomography with sample rotation: influence of a missing apple core in the recorded frequency space”

S. Vertu, *et al.*, *Centr. Eur. J. of Phys.* 7, p. 22 (2009)

Multiview Tomography



Multiview Tomography

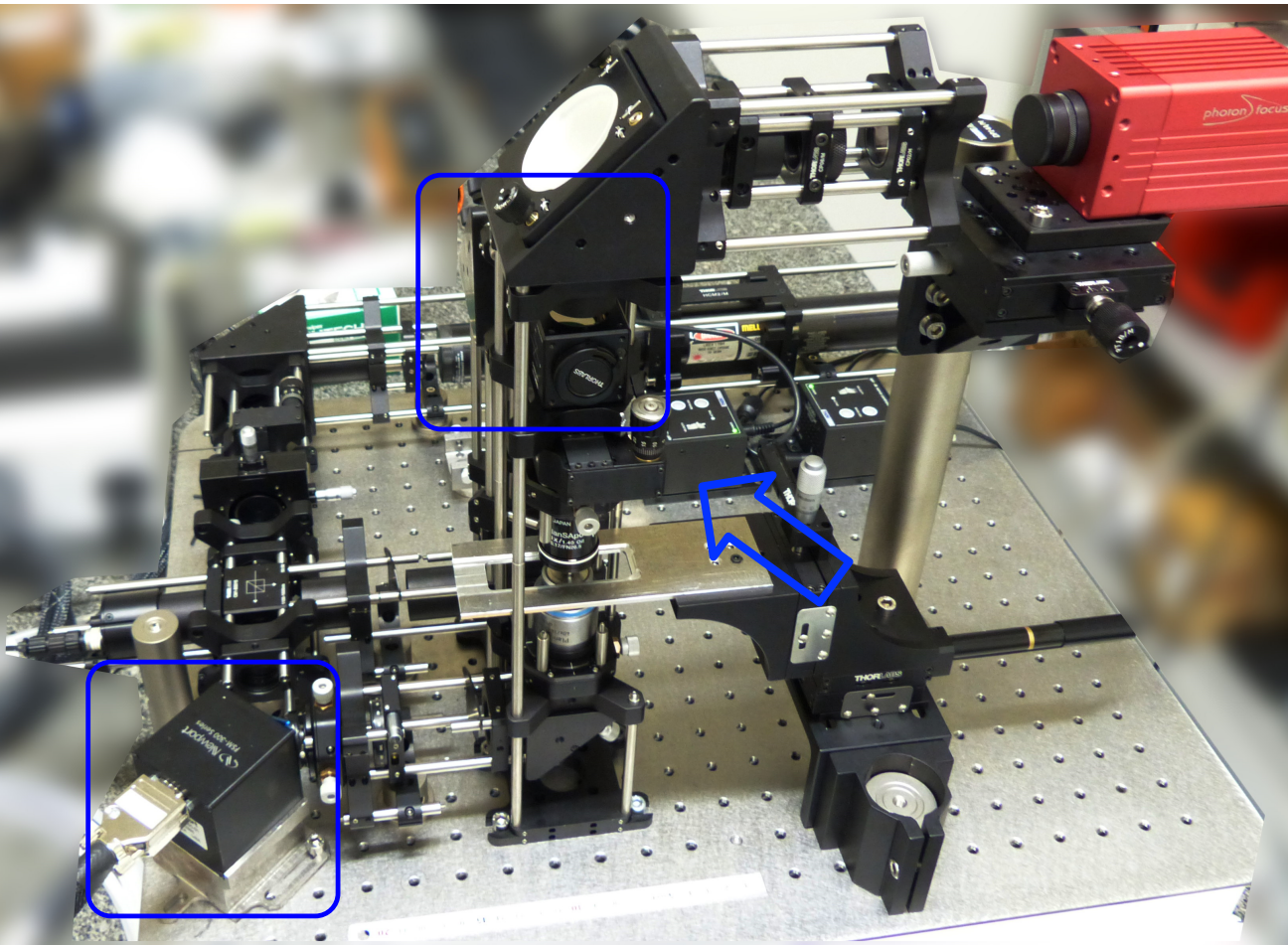


**Improved and isotropic resolution in tomographic diffractive microscopy
combining sample and illumination rotation**

S. Vertu, *et al.*, *Centr. Eur. J. of Phys.* **9**, p. 969 (2011)

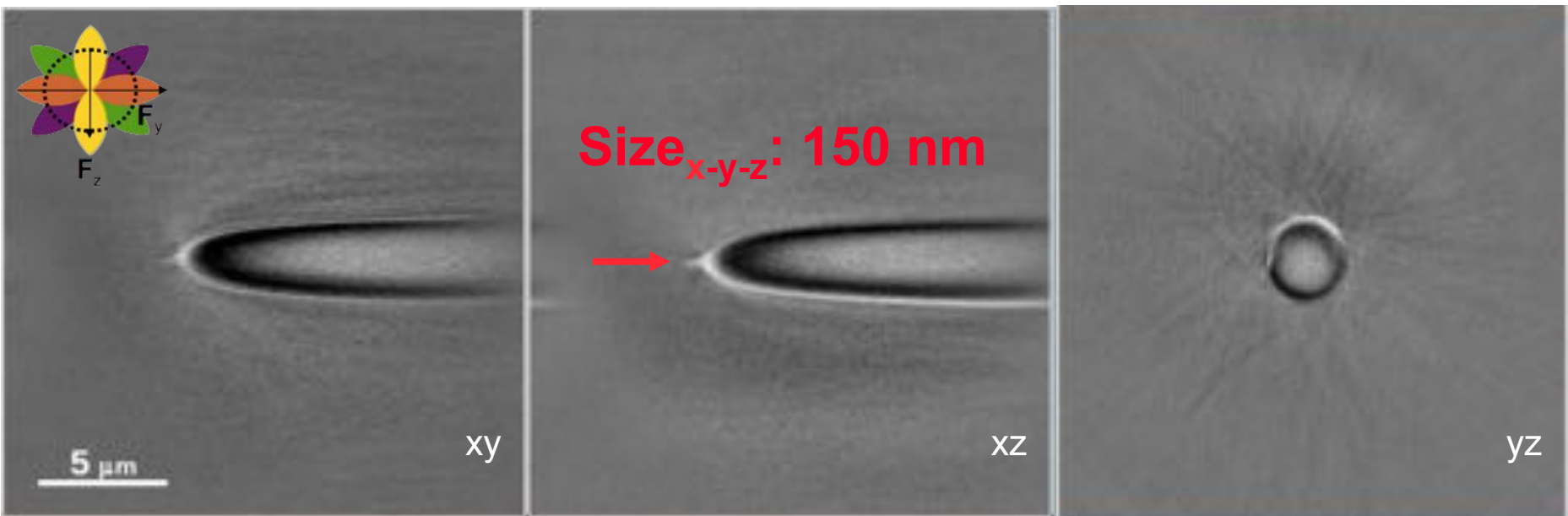
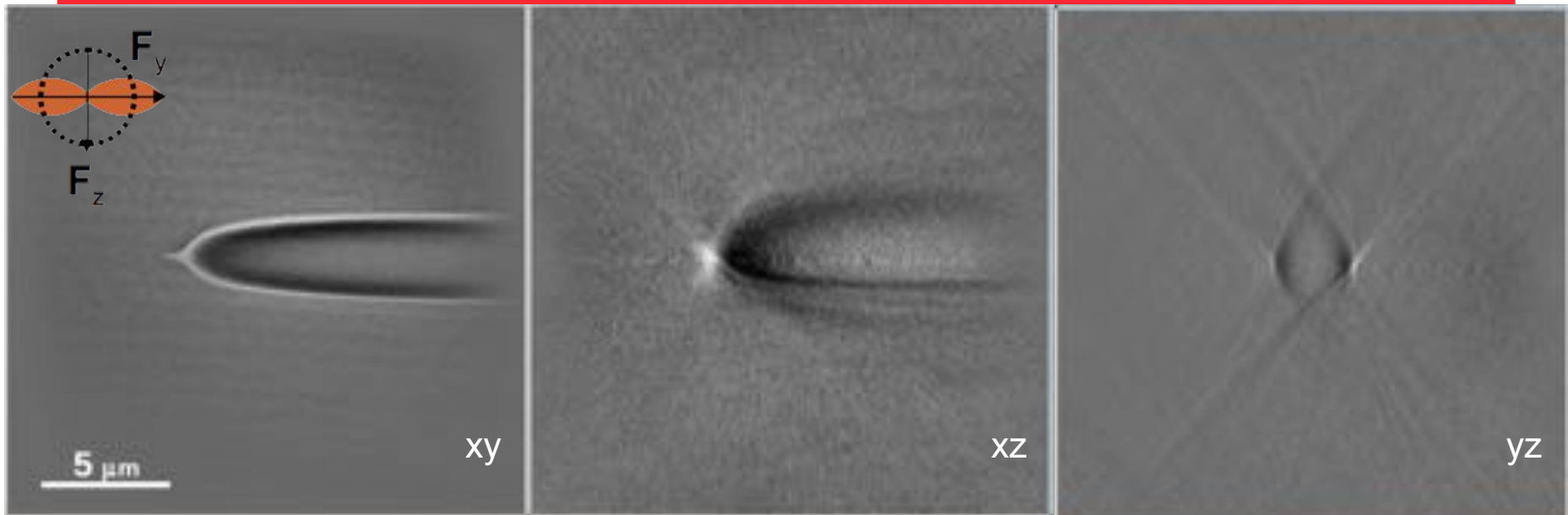
Towards High NA, IsoResolution

$\lambda=633\text{nm}$ or 475nm , $NA_{\text{obj}}=1.4$, $NA_{\text{cond}}=1.4$



Fast acquisition (less than 10s for 1 object orientation / 400 illuminations)
Real-time reconstruction for each object orientation (1 volume of data each 3s)₅₂

Optical Fiber Tip ($\lambda=475$ nm $R_{\text{predicted}}=95$ nm)



Zeolith microcrystal

5 μm

(a)

(b)

(c)

(d)

(e)

(a): 1 view

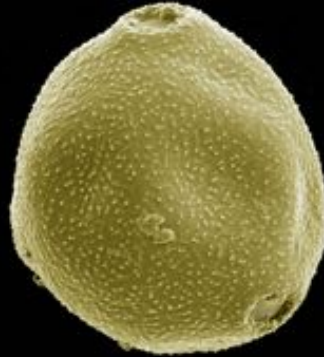
(b-e) fusion of 8 views

1 view

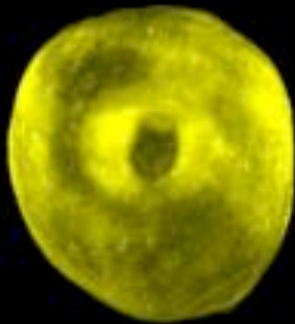
1 view deconvolved

fusion of 8 views

Betula Pollen



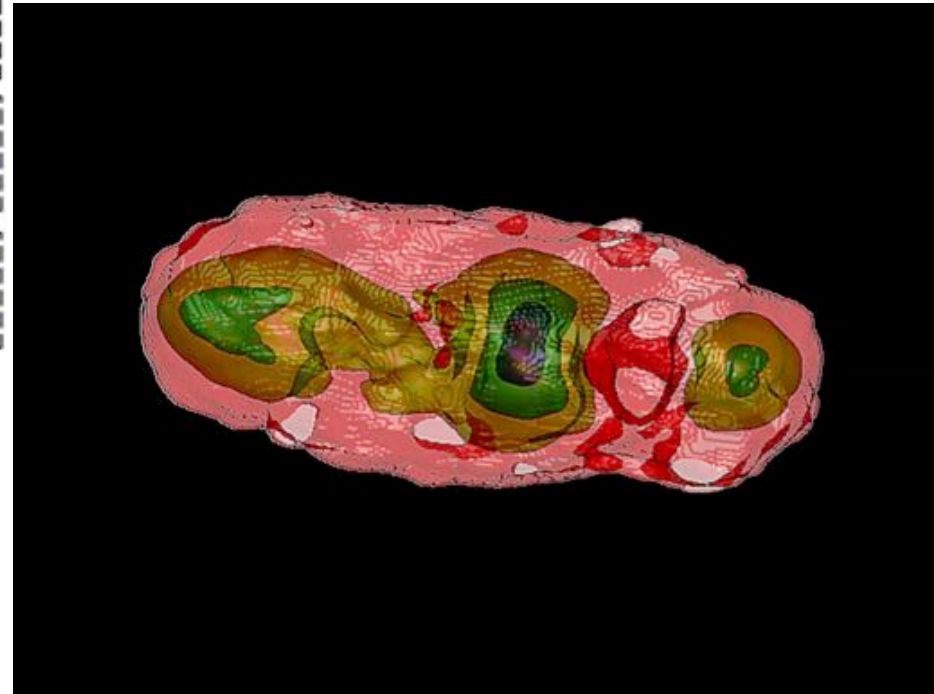
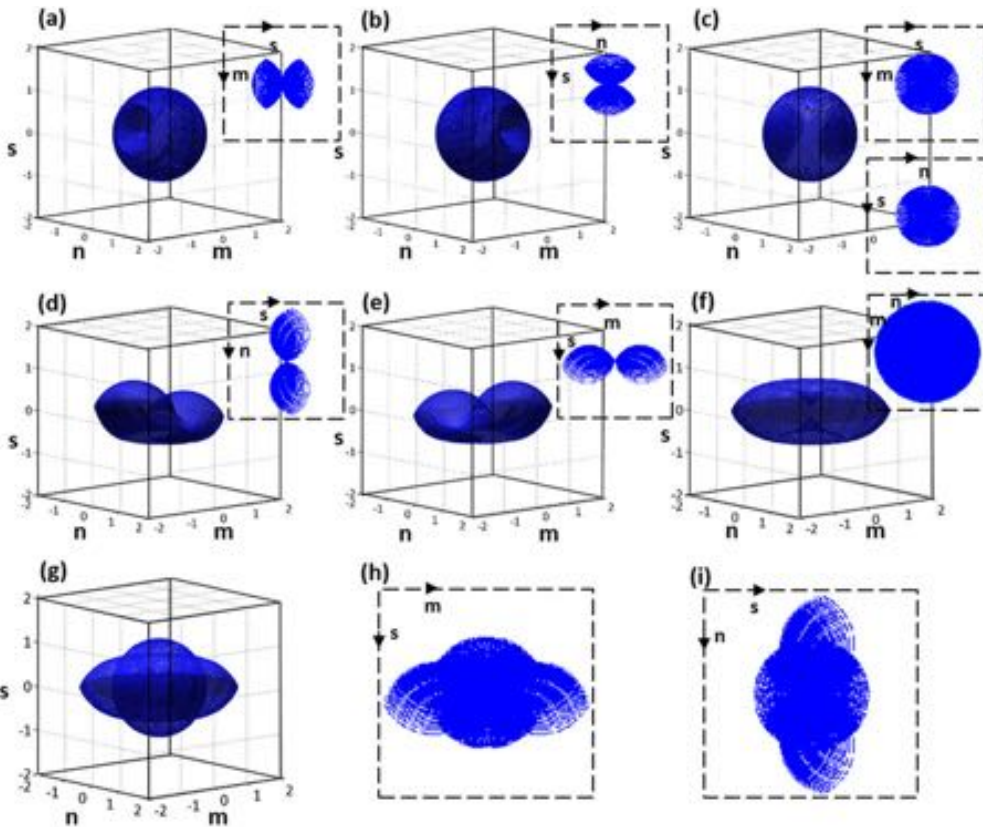
<http://www.vcbio.science.ru.nl>



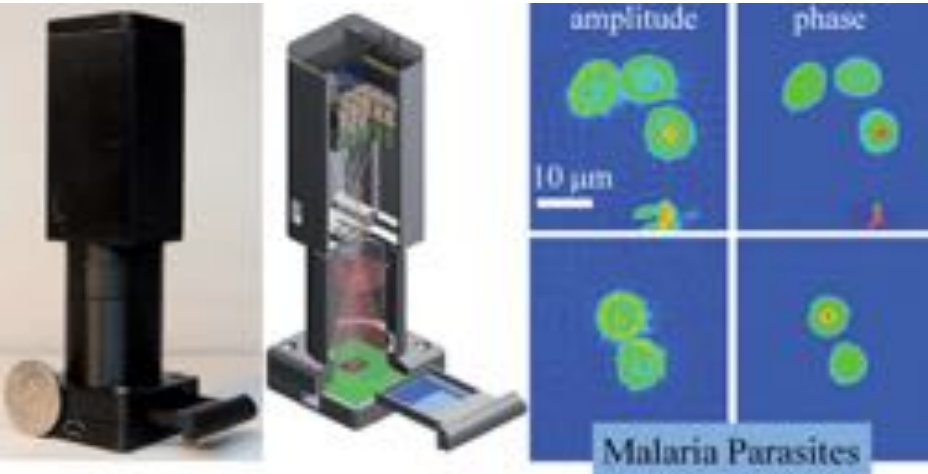
Betula Pollen

Integrated dual-tomography for refractive index analysis of free-floating single living cell with isotropic superresolution

B. Vinoth, *et al.*, Scientific Reports 8, 5943 (2018)



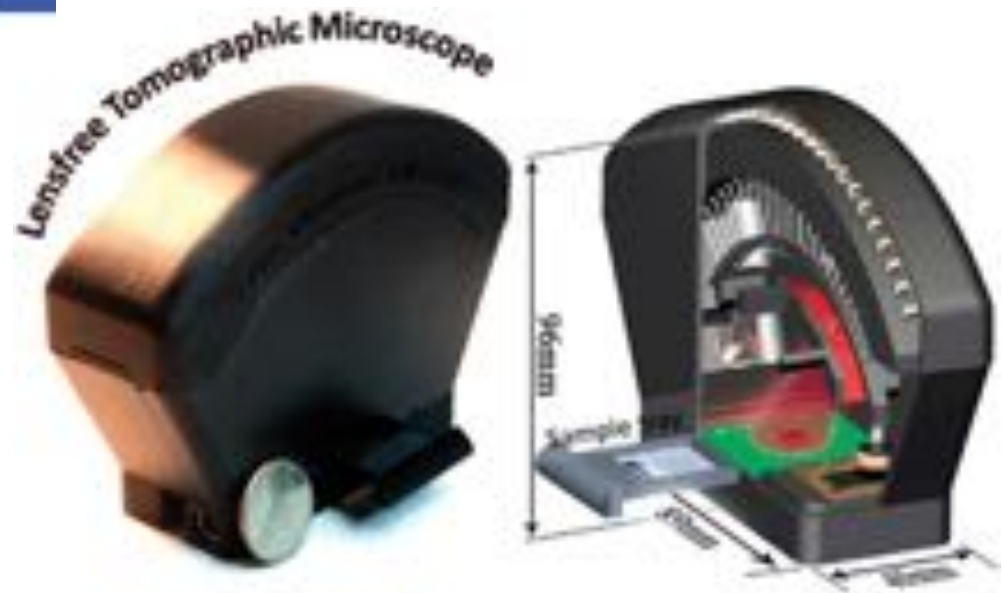
Low-cost microscopy/tomography



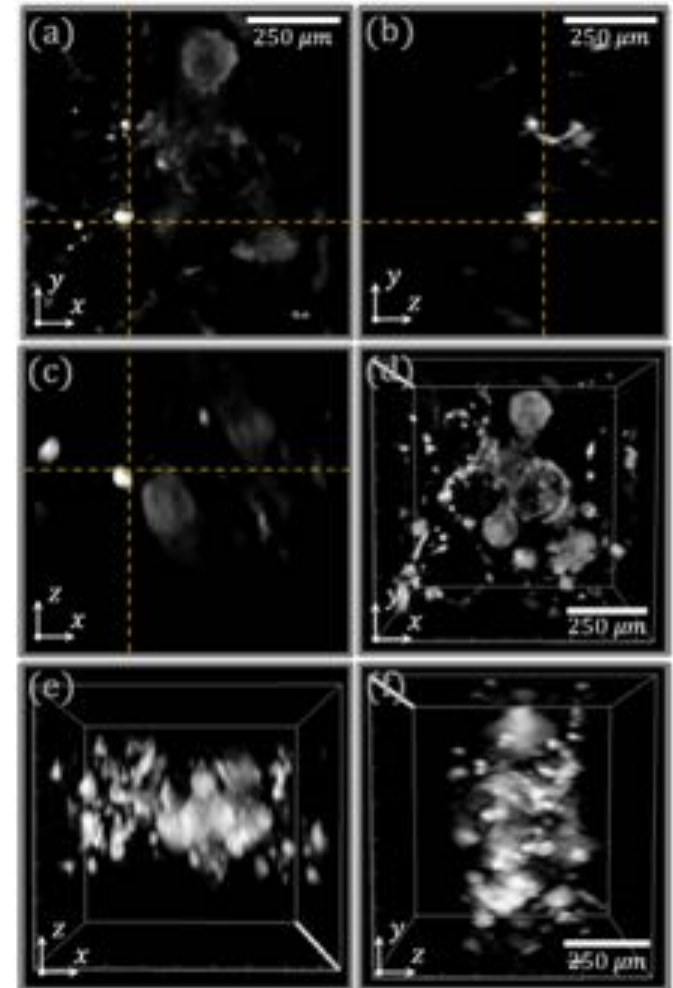
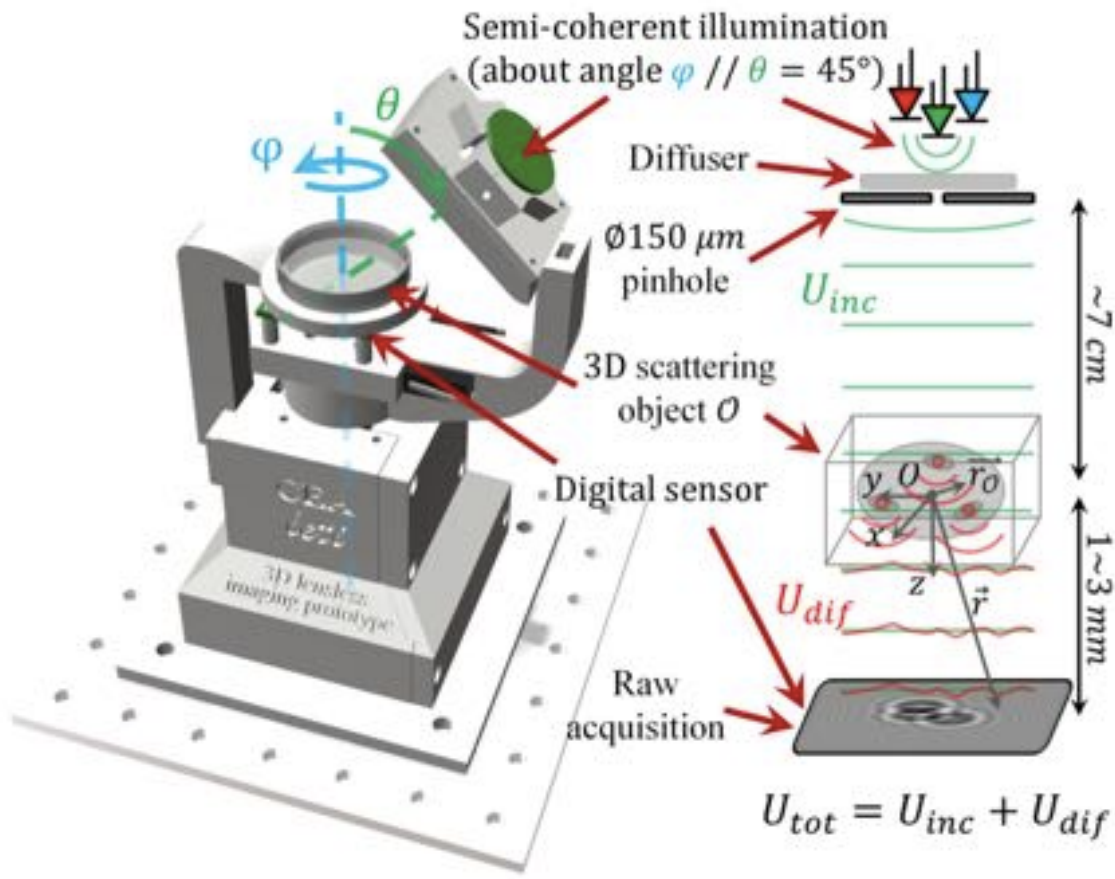
Aydogan Ozcan's group
UCLA

**Lens-free optical tomographic
microscope with a large imaging
volume on a chip**

S. O. Isikman, *et al.*,
PNAS 1015638108 (2011)



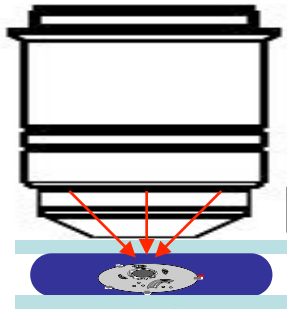
Low-cost microscopy/tomography



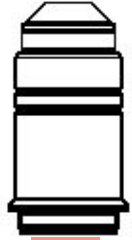
Comparative study of fully three-dimensional reconstruction algorithms for lens-free microscopy

A. Berdeu, *et al.*, *Appl. Opt.* **56**, p. 3939 (2017)

Drawback : Speed



Intensity Microscopy:
Transmission,
Phase contrast, DIC...

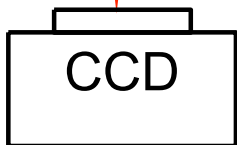


**Incoherent
Parallel**

⇒ **Ultrafast** 😊

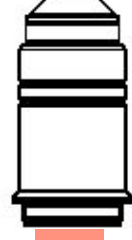
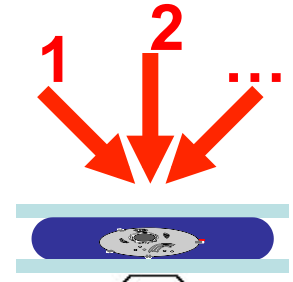


**Not-quantitative
Low-resolution**

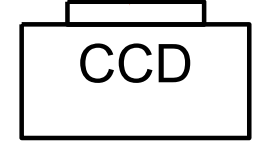


Diffractive
Tomographic
Microscopy

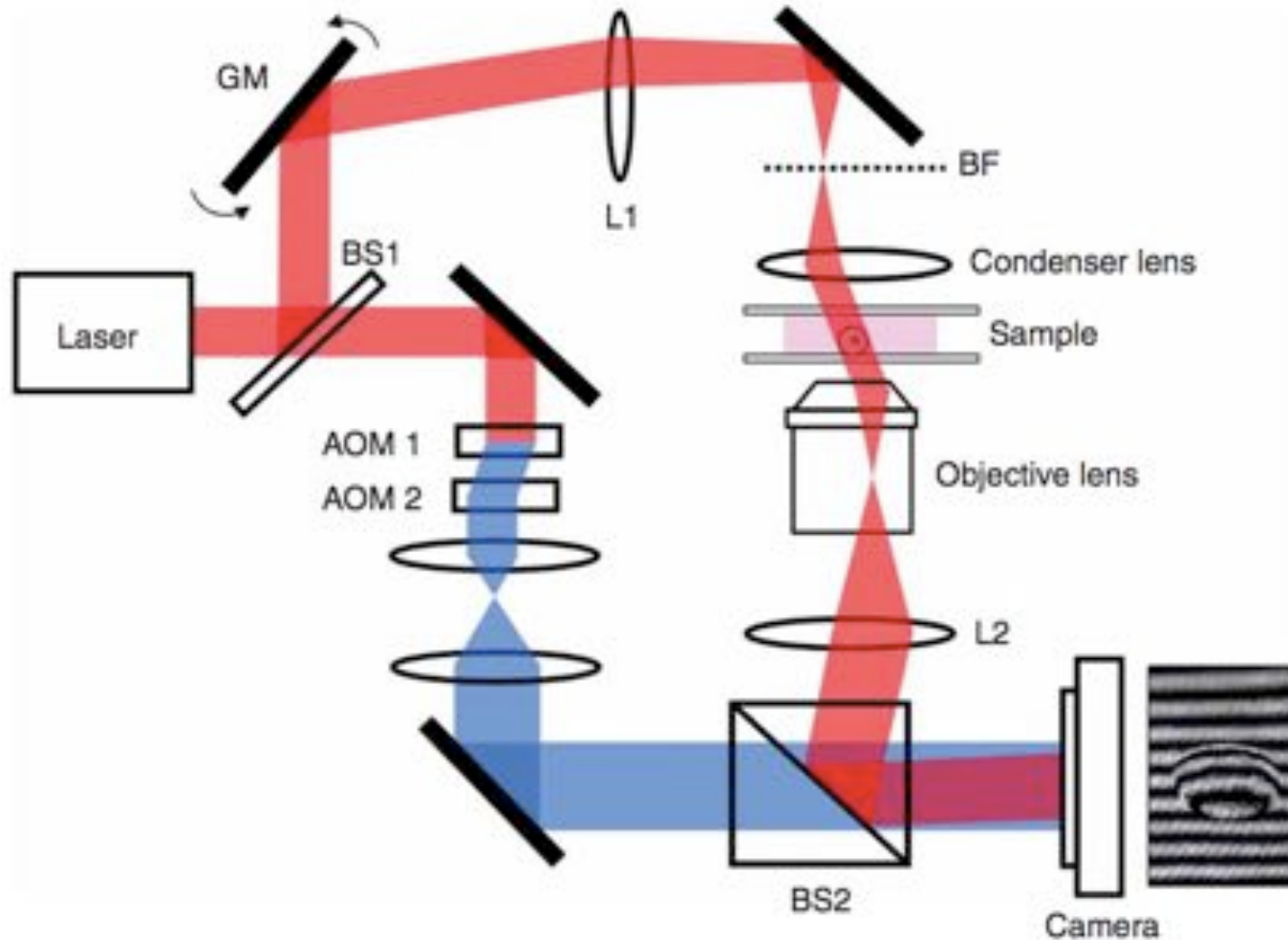
**Coherent
Sequential
⇒ Slow** ☹️



**Quantitative
High-resolution**



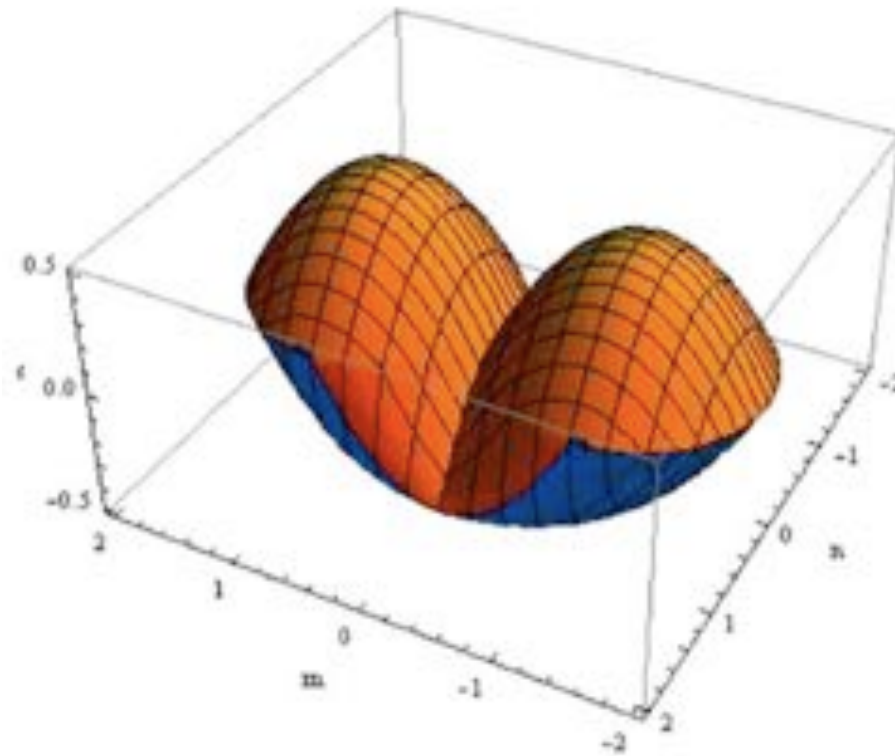
Fast 1-D Scanning



“Tomographic phase microscopy”

W. Choi, *et al.*, Nat. Meth. **4**, p. 717 (2007)

1-D Scanning



“The overall shape takes a form of what we might call a “peanut.”

“Image formation in holographic tomography”

S. Shan Kou, and C. J. R. Sheppard, *Opt. Lett.* **33**, p. 2362 (2008)

Acquisition Reconstruction Display

1.4 ms for 1 angle

Projection

GPU

Addition

GPU

Data preparation

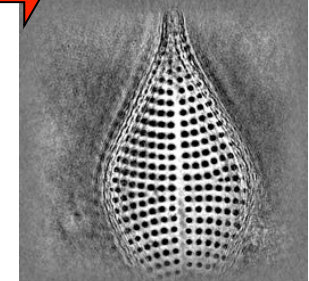
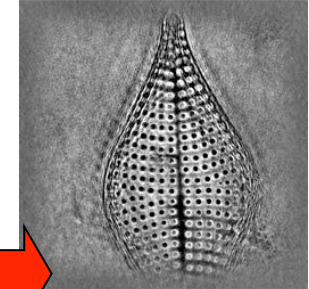
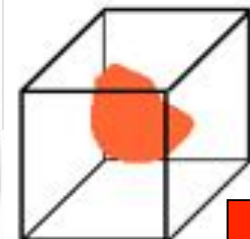
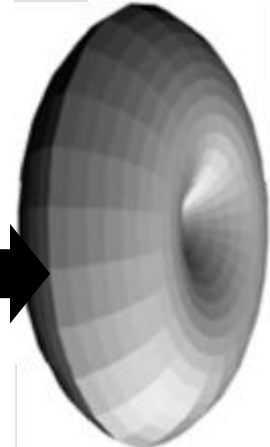
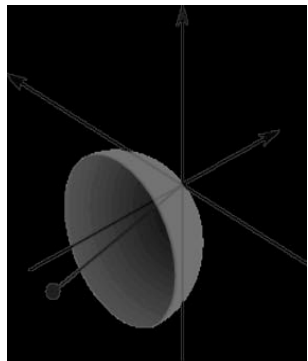
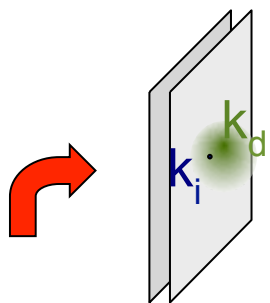
+ 3-D FT

GPU

240 ms

3-D
frequency
support

Display
(±1ms)



Off-axis
2-D FT

GPU

Object frequencies

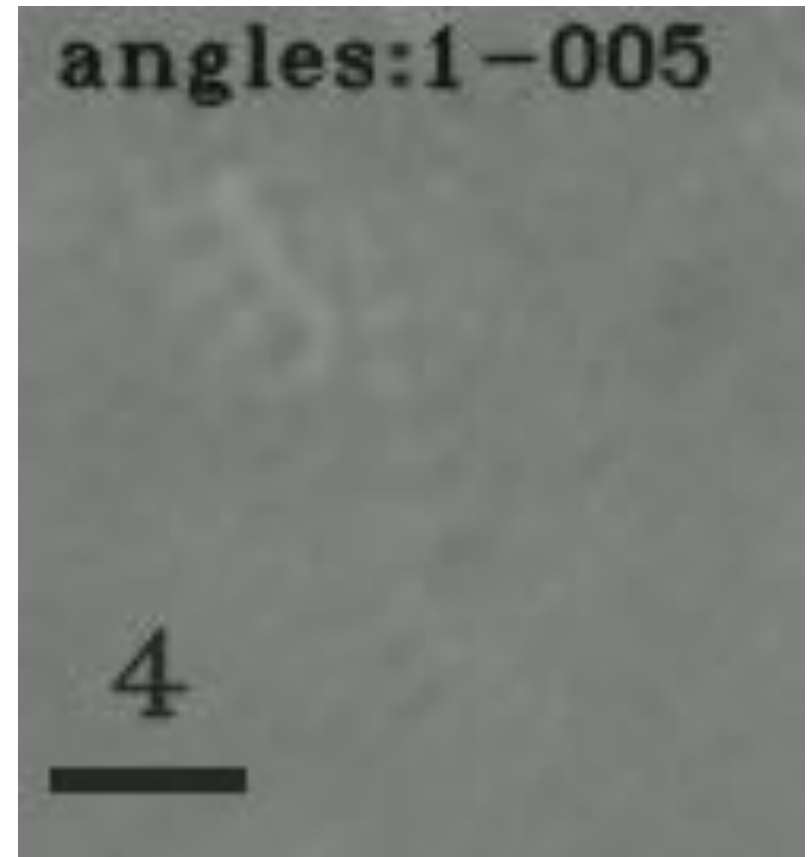
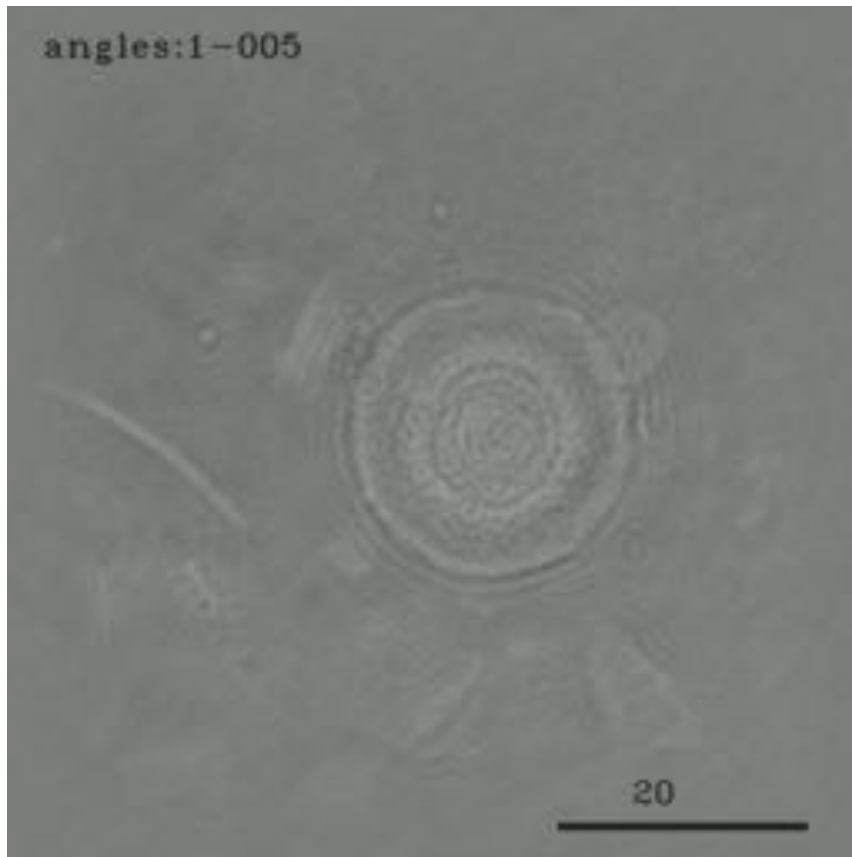
New angle

CPU->Mirror

1 hologram
Camera->CPU



GPU Reconstruction



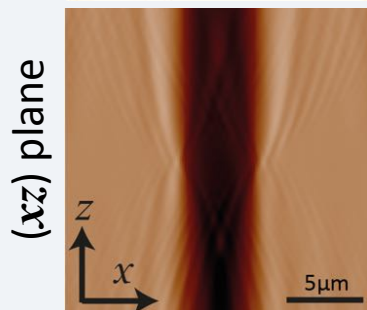
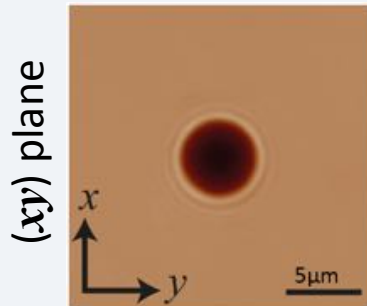
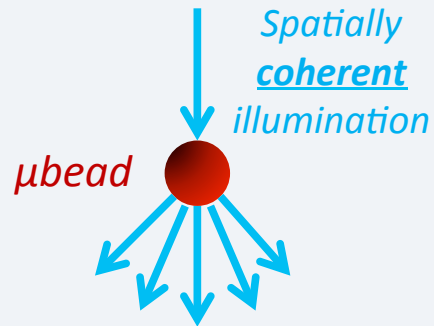
Nvidia Tesla C2075, Cuda, FFTW : 3.5 3D images/s

Tomographic diffractive microscopy: towards high-resolution 3-D real-time data acquisition, image reconstruction and display of unlabeled samples

J. Bailleul, *et al.*, Opt. Comm. **422**, p. 28 (2018)

Other possible approach

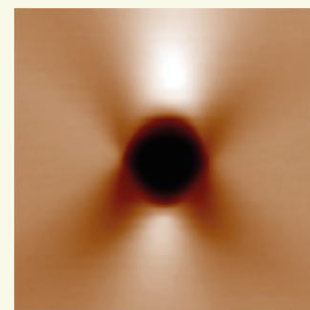
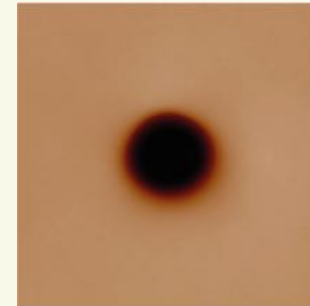
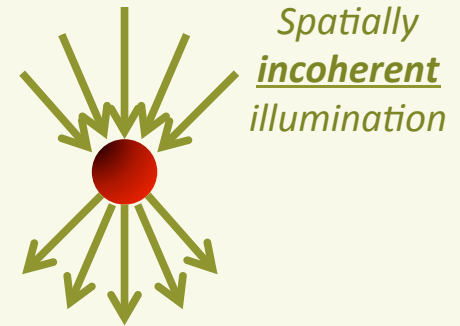
Conventional QPI



⊗ Limited axial resolution

New approach

Bon *et al.*,
Optics Express
2014

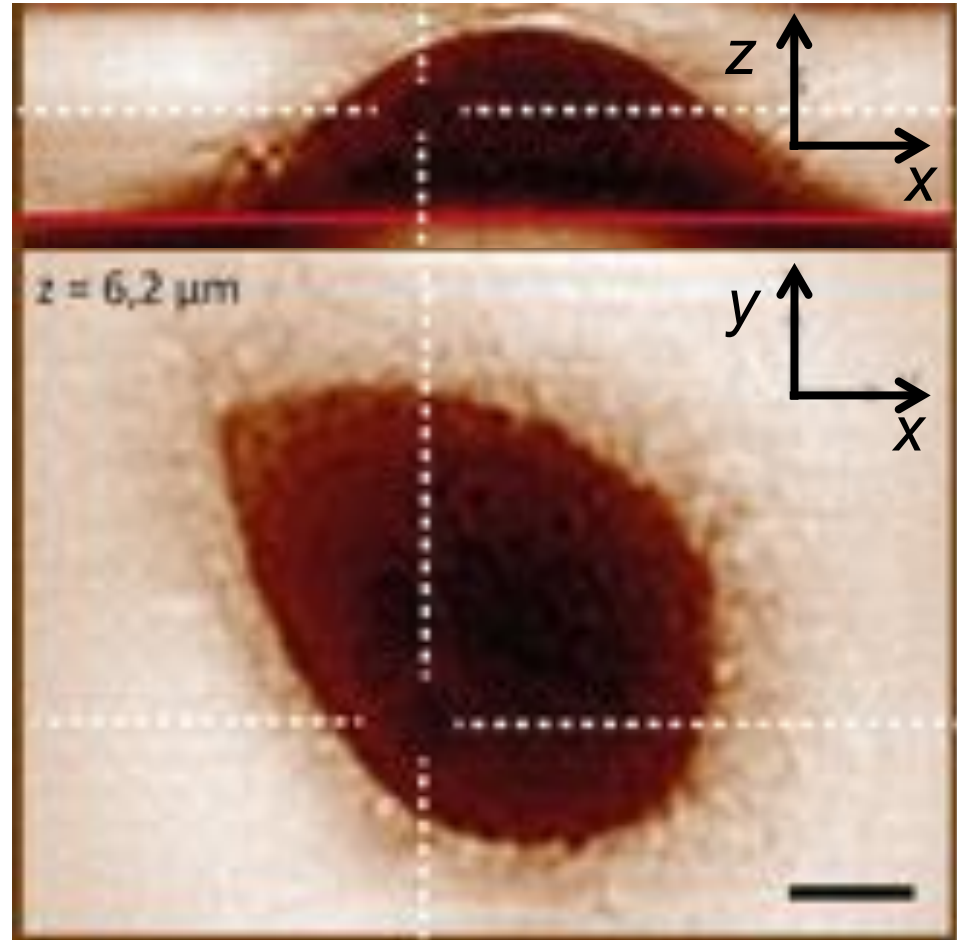
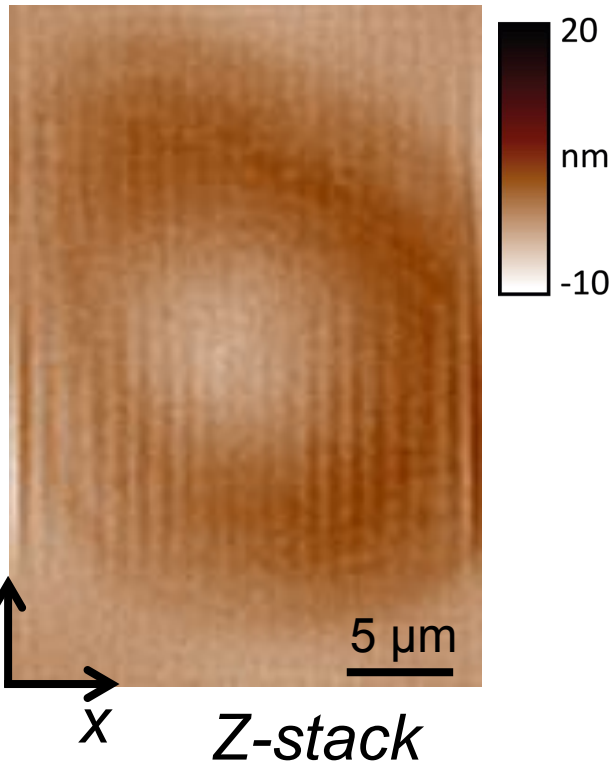


☺ 3D resolution \leftrightarrow fluorescence

3D live cell imaging

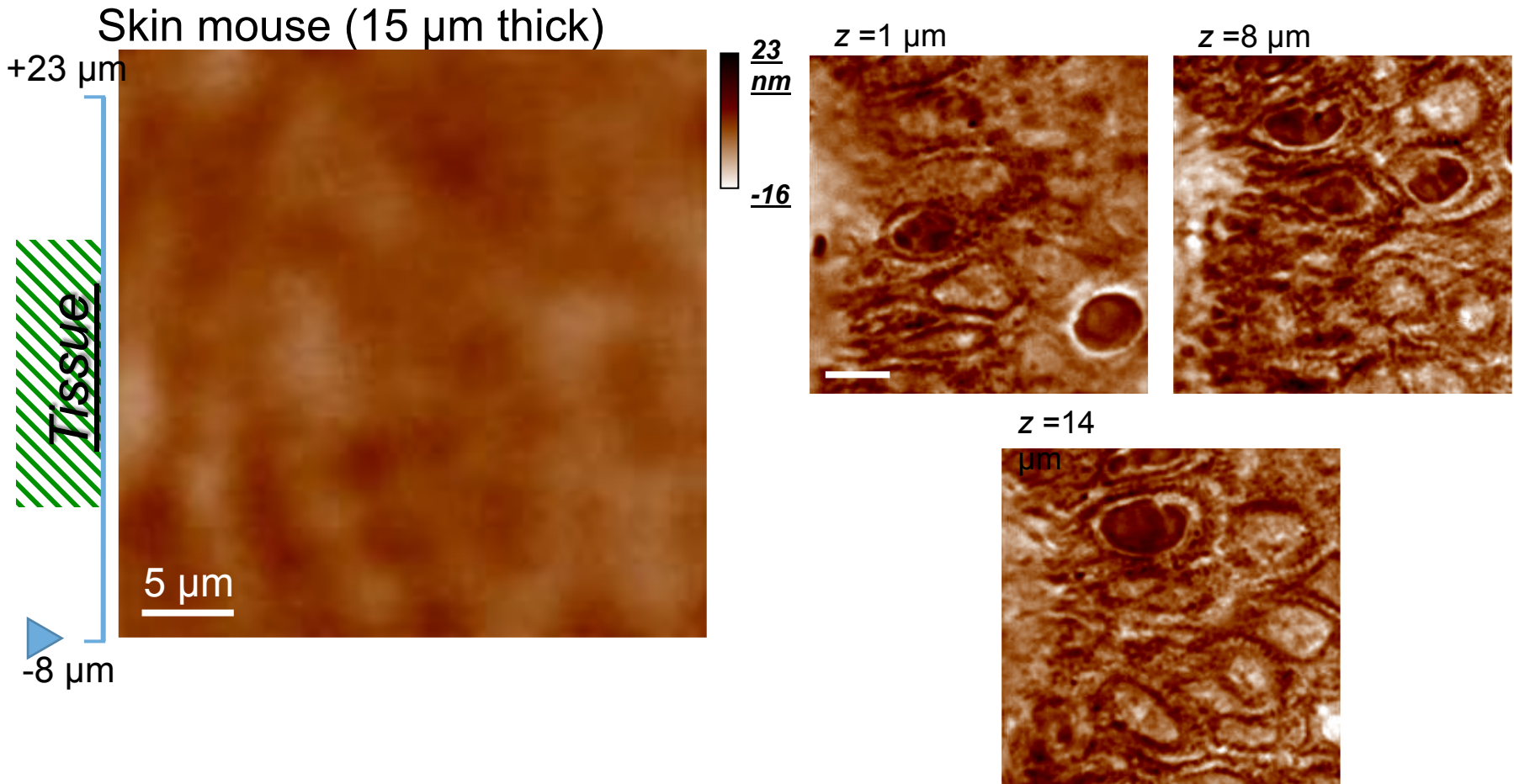
Living COS-7 cell

*OPD with spatially Incoh. Illum.
+ 3D deconvolution*



- ✓ **3D shape of the cell**
- ✓ **Fast acquisition, compatible with live imaging (just a z-stack!)**

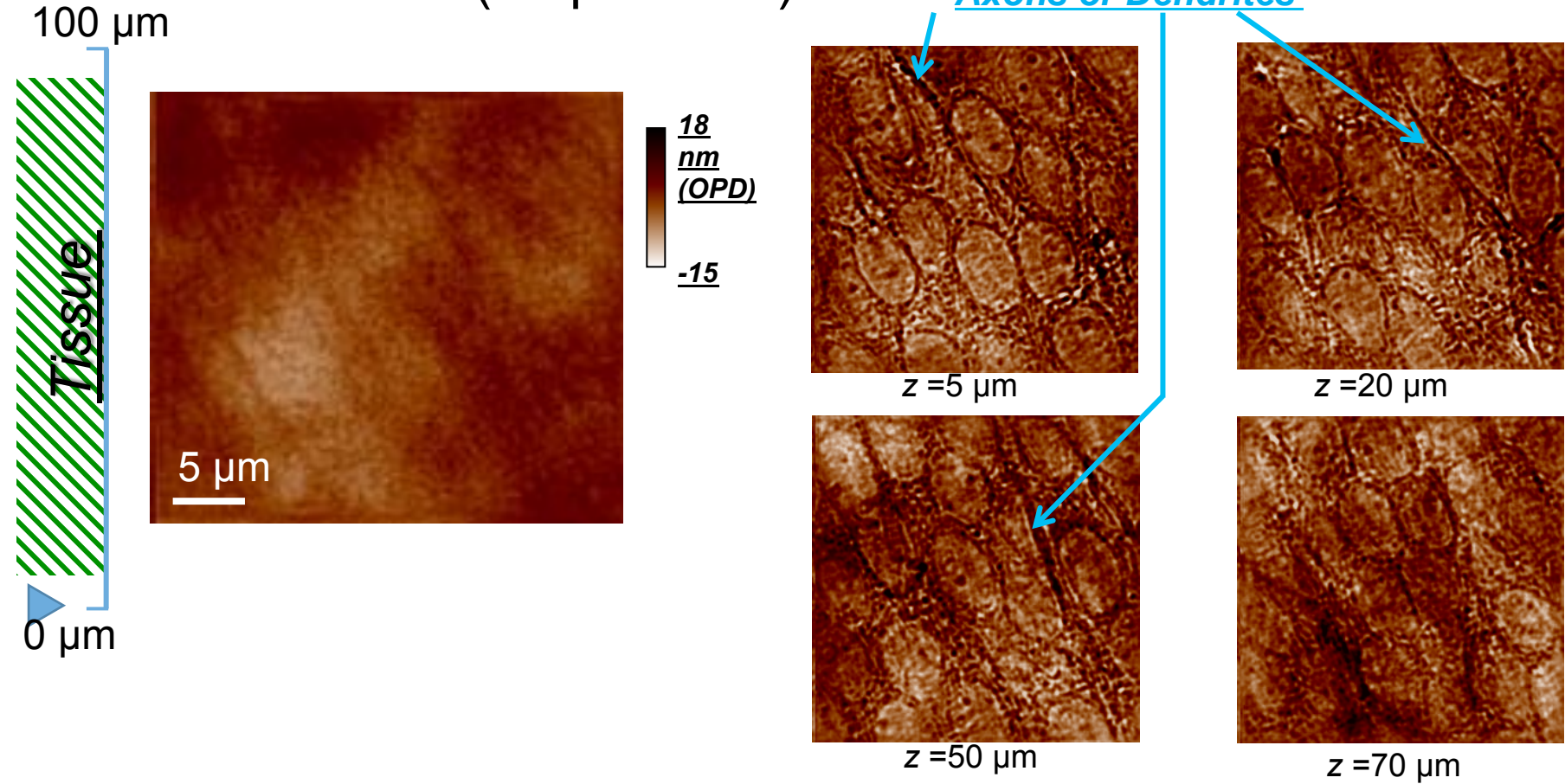
Fixed tissue imaging (1/2)



✓ Cell layer visualization without labeling

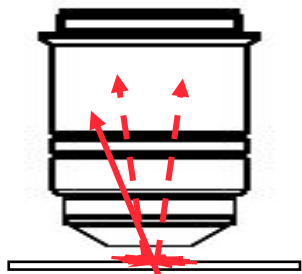
Fixed tissue imaging (2/2)

Brain slice (90 μm thick)

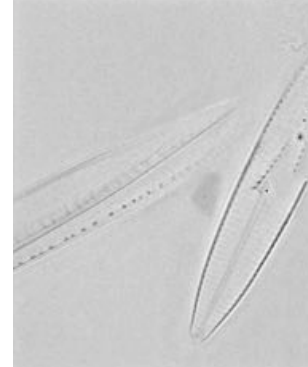


✓ Thin structures visible even after few tens of microns

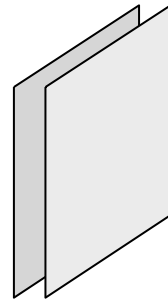
Multimode Imaging



→
Synthetic
reconstruction



Refraction
Microscopy

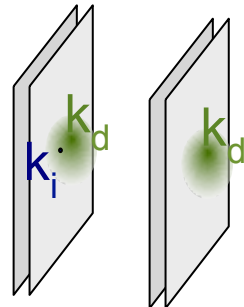
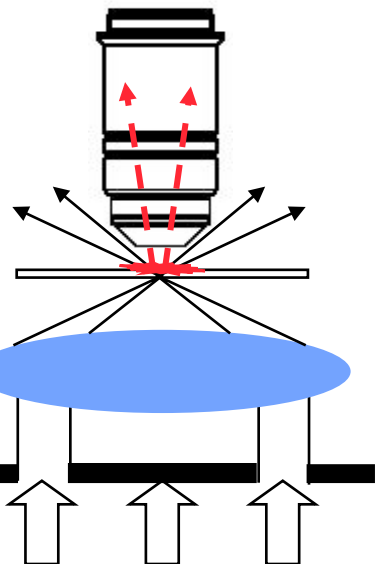


Wavefront

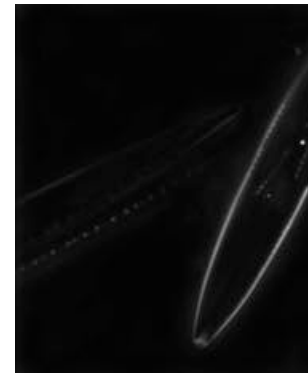
→
Compute
Intensity
(2)



Wide-Field
Microscopy



→
FT
(2)



High-NA
Dark-Field
Microscopy

Conclusion

Unprepared samples => use of a new kind of information $\langle n \rangle$

High resolution imaging: $\lambda/(3.5NA)$ lateral experimentally demonstrated

Challenges:

- RT acquisition/reconstruction/display**
- polarimetric TDM**
- “true” superresolution?**